

User manuals

NucleoSpin® Plant Pathogen

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Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned product are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols, can all be found on the product website and accessed easily via the QR code.

QR-Code product website



qr.mn-net.com/qr/(241)740170

Use the following QR code or the link below for direct access to the user manual.

QR-Code user manual



qr.mn-net.com/qr/(IFU)740170

It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This leaflet does not replace the full manual!



Protocol at a glance (Rev. 01) NucleoSpin® Plant Pathogen - Viral RNA and DNA isolation from plant samples

1 Homogenize and lyse sample Popular PFL All Light PFR Mix 56 °C, 5 min 14,000 x g, 1 min Option B: Bead Beating Option B: Bead It 4,000 xg, I min It 4,000 xg, I min			
Sample		Option A: Crush	300 μL PFL
Mix 56 °C, 5 min 14,000 x g, 1 min Option B: Bead Beating 300 μL PFL 12 μL PFR Agitate 2 x 30 s 56 °C, 5 min Remove steel balls 14,000 x g, 1 min Load lysate 14,000 x g, 30 s 2 Adjust binding conditions Bind RNA and DNA Load 650 μL sample 14,000 x g, 30 s 4 Wash silica membrane 1st wash 2nd wash 500 μL PFW1 14,000 x g, 1 min 2nd wash 500 μL PFW2 14,000 x g, 1 min			12 µL PFR
Option B: Bead Beating Option B: Bead Beating 300 μL PFL 12 μL PFR Agitate 2 x 30 s 56 °C, 5 min Remove steel balls 14,000 x g, 1 min Load lysate 14,000 x g, 30 s 2 Adjust binding conditions 3 Bind RNA and DNA Load 650 μL sample 14,000 x g, 30 s Load 650 μL sample 14,000 x g, 30 s 4 Wash silica membrane 14,000 x g, 1 min 500 μL PFW2 14,000 x g, 1 min 500 μL RNase-free H ₂ O RT, 1 min 14,000 x g,	Junpic	\bigcirc	Mix
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Option B: Bead Beating 300 µL PFL 12 µL PFR Agitate 2 x 30 s 56 °C, 5 min Remove steel balls 14,000 x g, 1 min Load lysate 14,000 x g, 30 s 300 µL PFB Mix RT, 5 min 3 Bind RNA and DNA Load 650 µL sample 14,000 x g, 30 s 4 Wash silica membrane 1st wash 2nd wash 500 µL PFW2 14,000 x g, 1 min 500 µL RNase-free H₂O RT, 1 min 14,000 x g,			
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Remove steel balls			Agitate 2 x 30 s
14,000 x g, 1 min Load lysate 14,000 x g, 30 s 2 Adjust binding conditions 3 Bind RNA and DNA Load 650 μL sample 14,000 x g, 30 s 4 Wash silica membrane 1st wash 2nd wash 2nd wash 3rd wash 500 μL PFW2 14,000 x g, 1 min 50 μL RNase-free H ₂ O RT, 1 min 14,000 x g, RT, 1 min 14,000 x g,			56 °C, 5 min
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3 rd wash 500 μL PFW2 14,000 × g, 1 min 5 Elute RNA and DNA 50 μL RNase-free H ₂ O RT, 1 min 14,000 × g,			
14,000 x g, 1 min 5 Elute RNA and DNA 50 μL RNase-free H ₂ O RT, 1 min 14,000 x g,			1 min
5 Elute RNA and DNA 50 μ L RNase-free H ₂ O RT, 1 min 14,000 \times g ,	3 rd wash	<i>≥</i> 5	
5 Elute RNA and DNA 50 μL RNase-free H ₂ O RT, 1 min 14,000 × g,			
and DNA 50 µL RNase-free H ₂ O RT, 1 min 14,000 x g,	5 Fluto DNA	25	1 171111
RT, 1 min 14,000 x g,			50 μL RNase-free H ₂ O
-			·
1 min		\\	14,000 × <i>g</i> ,
		<u></u>	1 min



QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:



www.mn-net.com/sds



We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact

our Technical Support.

Contact MN

Germany and international

MACHEREY-NAGEL GmbH & Co. KG

Valencienner Str. 11 · 52355 Düren · Germany

Tel.: +49 24 21 969-0

Toll-free: 0800 26 16 000 (Germany only) E-mail: info@mn-net.com

support@mn-net.com

Technical support Bioanalysis
Tel.: +49 24 21 969-333

E-mail: USA

MACHEREY-NAGEL Inc.

924 Marcon Blvd. · Suite 102 · Allentown PA, 18109 · USA

Toll-free: 888 321 6224 (MACH) E-mail: sales-us@mn-net.com

France

MACHEREY-NAGEL SAS

1, rue Gutenberg – BP135 \cdot 67720 Hoerdt Cedex \cdot France

Tel.: +33 388 68 22 68 E-mail: sales-fr@mn-net.com

MACHEREY-NAGEL SAS (Société par Actions Simplifiée) au capital de

186600 €

Siret 379 859 531 00020 · RCS Strasbourg B379859531

N° intracommunautaire FR04 379 859 531

Switzerland

MACHEREY-NAGEL AG

Hirsackerstr. 7 \cdot 4702 Oensingen \cdot Switzerland

Tel.: +41 62 388 55 00
E-mail: sales-ch@mn-net.com

