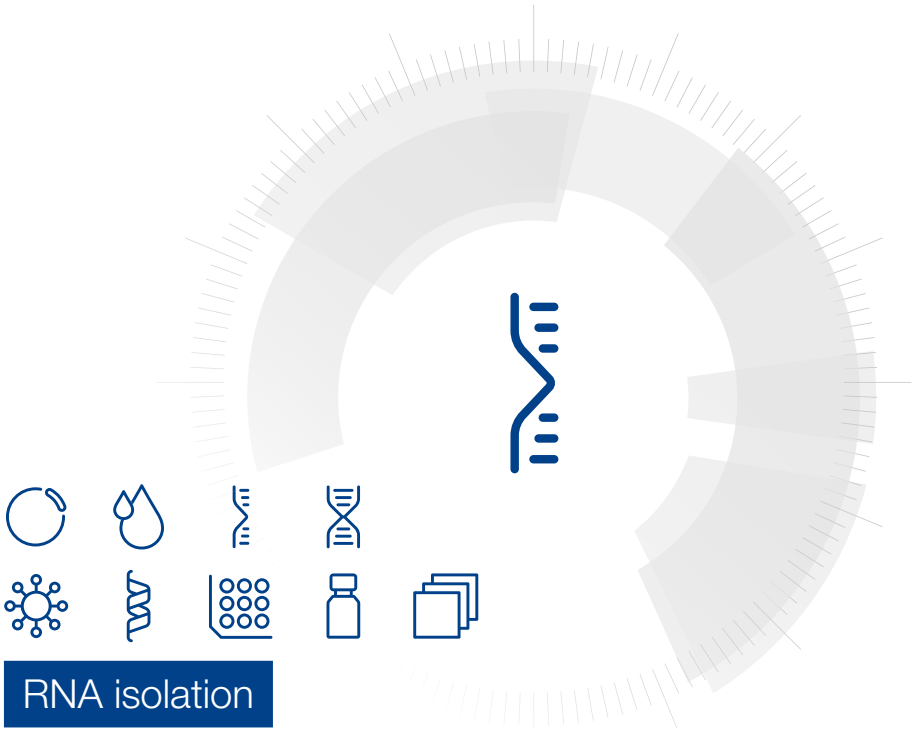
Bioanalysis

MACHEREY-NAGEL

User manual



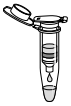
RNA isolation

- NucleoSpin® RNA XS

May 2023 / Rev. 12

RNA isolation

Protocol at a glance (Rev. 12)

		XS NucleoSpin® RNA XS
1	**Supply sample**	Use up to 10^5 cultured cells or 5 mg tissue samples
2	**Lyse and homogenize cells**	 100 µL RA1 2 µL TCEP Mix
3	**Add Carrier RNA**	 5 µL Carrier RNA working solution Mix
4	**Filtrate lysate** *(optional)*	  11,000 x *g*, 30 s
5	**Adjust RNA binding conditions**	100 µL 70 % ethanol Mix
6	**Bind RNA**	  Load lysate 11,000 x *g*, 30 s
7	**Desalt silica membrane**	  100 µL MDB 11,000 x *g*, 30 s
8	**Digest DNA**	 25 µL DNase reaction mixture RT, 15 min
9	**Wash and dry silica membrane**	 1st wash: 100 µL RA2 2nd wash: 400 µL RA3 3rd wash: 200 µL RA3 1st and 2nd  11,000 x *g*, 30 s 3rd  11,000 x *g*, 2 min
9	**Elute highly pure RNA**	  10 µL RNase-free H_2O 11,000 x *g*, 30 s

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1 Components

1.1 Kit contents

	NucleoSpin® RNA XS		
REF	**10 preps 740902.10**	**50 preps 740902.50**	**250 preps 740902.250**
Lysis Buffer RA1	6 mL	25 mL	125 mL
Wash Buffer RA2	2 × 1 mL	15 mL	2 × 15 mL
Wash Buffer RA3 (Concentrate)*	6 mL	12 mL	50 mL
Membrane Desalting Buffer MDB	10 mL	10 mL	50 mL
Reaction Buffer for rDNase	7 mL	7 mL	30 mL
rDNase, RNase-free (lyphilized)*	1 vial (size A)	1 vial (size C)	2 vials (size D)
Carrier RNA*	300 µg	300 µg	300 µg
Reducing Agent TCEP*	14 mg	3 × 14 mg	2 × 107 mg
RNase-free H_2O	13 mL	13 mL	13 mL
NucleoSpin® Filters (violet rings)	10	50	250
NucleoSpin® RNA XS Columns (light blue rings – plus Collection Tubes)	10	50	250
Collection Tubes (2 mL)	30	150	750
Collection Tubes (1.5 mL)	10	50	250
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* For preparation of working solutions and storage conditions see section 3.

1.2 Reagents, consumables, and equipment to be supplied by user

Reagents

- 96 – 100 % ethanol (to prepare Wash Buffer RA3 and for the clean up procedure, section 5.3)
- 70 % ethanol (to adjust RNA binding condition)

Consumables

- 1.5 mL microcentrifuge tubes
- Sterile RNase-free tips

Equipment

- Manual pipettors
- Centrifuge for microcentrifuge tubes
- Vortex mixer
- Personal protection equipment (lab coat, gloves, goggles)

1.3 About this user manual

It is strongly recommended reading the detailed protocol sections of this user manual if the **NucleoSpin® RNA XS** kit is used for the first time. Experienced users, however, may refer to the Protocol at a glance instead. The Protocol at a glance is designed to be used only as a supplemental tool for quick referencing while performing the purification procedure.

All technical literature is available on the internet at www.mn-net.com.

Please contact Technical Service regarding information about changes of the current user manual compared to previous revisions.

2 Product description

2.1 The basic principle

One of the most important aspects isolating RNA is to prevent degradation of the RNA during the isolation procedure. With the **NucleoSpin® RNA** methods, cells are lysed by incubation in a solution containing large amounts of chaotropic ions. This lysis buffer immediately inactivates RNases – which are present in virtually all biological materials – and creates appropriate binding conditions which favor adsorption of RNA to the silica membrane. Contaminating DNA, which is also bound to the silica membrane, is removed by an rDNase solution which is directly applied onto the silica membrane during the preparation (RNase-free rDNase is supplied with the kit). Simple washing steps with two different buffers remove salts, metabolites and macromolecular cellular components. Pure RNA is finally eluted under low ionic strength conditions with RNase-free H_2O (supplied).

The RNA preparation using **NucleoSpin® RNA** kits can be performed at room temperature. The eluate, however, should be treated with care because RNA is very sensitive to trace contaminations of RNases, often found on general lab ware, fingerprints and dust. To ensure RNA stability, keep RNA frozen at -20 °C for short term or -70 °C for long term storage.

2.2 Kit specifications

- The **NucleoSpin® RNA XS** kit is recommended for the isolation of RNA from very small samples. Typical sample material comprises small amounts of cells (up to 1×10^5) and tissue (up to 5 mg) such as pellets of cultured cells, laser-captured cells, microdissected cryosections, biopsy samples, fine needle aspirates, and flow cytometer sorted cells (Table 1, page 8).
- The innovative column design with a funnel shaped thrust ring and a small silica membrane area allows elution of RNA in as little as 5 – 30 µL. Thus, **highly concentrated RNA** is eluted, ready for common downstream applications (e.g., RT-PCR).
- The **RNA yield** strongly depends on the sample type, quality, and amount (see Table 2, page 8 for details).
- **High quality RNA** (RNA Integrity Number (RIN) > 9 according to Agilent 2100 Bioanalyzer assays) can be obtained from small samples (e.g., 10^3 cells, 0.1 mg tissue) as well as from larger samples (10^5 cells, 5 mg tissue). rRNA ratios (28S / 18S) of 1.8 – 2.0 can be obtained. Since RNA quality always depends on the sample quality, see section 6.3 for further aspects.
- The **NucleoSpin® RNA XS** kit allows purification of RNA with an A_{260}/A_{280} ratio generally exceeding 1.9 (measured in TE buffer pH 7.5). Due to the high RNA **purity** large amounts of eluates can be used as template in RT-PCR without inhibition (e.g., 8 µL of 10 µL eluates as template in a 20 µL qRT-PCR setup generating stronger signal compared

to reactions with less template in a LightCycler PCR with the Sigma SYBR Green Quantitative RT-PCR Kit).

- The **preparation time** is approximately 45 min for 12 samples.
- As **Reducing Agent TCEP** (Tris(2-carboxyethyl)phosphine) is supplied in the kit. TCEP is odorless, more stable, more specific for disulfide-bonds, and less toxic than other commonly used reducing agents.
- **Carrier RNA** (poly(-A) RNA: poly(A) potassium salt, prepared from ADP with polynucleotide phosphorylase) is included for optimal performance with smallest samples.

It is recommended adding Carrier RNA to the sample lysate (20 ng per sample). Such small amounts typically do not interfere with subsequent RT-PCR, even in oligo-dT primed reverse transcriptions. The small amount of Carrier RNA transfered into a reverse transcription reaction is commonly not significantly influencing the outcome of the reaction, due to the large excess of oligo-dT primer. The benefit of adding Carrier RNA to the sample lysate depends on sample type, amount, and kind of downstream RNA analysis. Carrier RNA should be omitted when subsequent to RNA isolation
- a poly-A RNA isolation is performed
- RNA sequencing is performed.

- **rDNase** is supplied in the kit. DNA contaminations are removed by on column digestion with rDNase. For most demanding applications (e.g., expression analysis of plasmid transfected cells, plastidial or mitochondrial genes) a subsequent digestion with rDNase in the eluate is possible.
- For research use only

Table 1: Kit specifications at a glance

Parameter	NucleoSpin® RNA XS
Format	Mini spin column – XS design
Sample material	Small amounts of tissue < 5 mg tissue, < 100 000 cultured cells
Fragment size	> 200 nt
Typical yield	See Table 2 for examples
A_{260}/A_{280}	1.9 – 2.1
Typical RIN (RNA integrity number)	> 9 (depending on sample quality)
Elution volume	5 – 30 µL
Preparation time	35 min/6 preps
Binding capacity	110 µg

Table 2: Overview on average yields of RNA isolation using NucleoSpin® RNA XS

Sample	Average yield
10^5 HeLa cells	1000 – 1500 ng
10^4 HeLa cells	100 – 150 ng
10^3 HeLa cells	10 – 15 ng
10^2 HeLa cells	0.1 – 1.5 ng
5 mg mouse kidney	5 – 8 µg
1 mg mouse kidney	2 µg

2.3 Handling, preparation, and storage of starting materials

Work environment

Maintain an RNase-free work environment. Wear gloves at all times during the preparation. Change gloves frequently

Sample storage and RNase inhibition

RNases can rapidly degrade RNA within the samples if samples are not protected from RNase activity after harvest. Use one of the following methods to avoid RNA degradation:

- Use freshly harvested sample for immediate lysis and RNA purification.
- Submerge and store samples in NucleoProtect® RNA or similar stabilization solutions. Make sure to allow for complete permeation of the sample with the stabilization solution before freezing it. Remove excess stabilization solution from the sample prior to RNA isolation according to the stabilization solution user manual.
- Flash freeze sample in liquid N2 immediately upon harvest and store at -70 °C. Frozen samples are stable up to 6 months. Mortar and pestle can be used to pulverize the sample in a frozen state. Make sure that the sample does not thaw prior to contact with lysis buffer.
- Store samples in Lysis Buffer RA1 after disruption at -70 °C for up to one year, at 4 °C for up to 24 hours or at room temperature for up to several hours. Samples frozen in Lysis Buffer RA1 should be thawed slowly before starting with the isolation of RNA.

Disruption and homogenization of sample material

- **Cultured cells in suspension:**
  Collect cells by centrifugation, remove supernatant and immediately add Lysis Buffer RA1 according to step 2 of the standard protocol (see sections 5.1, 6.1).
- **Adherent cell cultures (lysis in culture dish):**
  Completely aspirate cell-culture medium. Immediately add Lysis Buffer RA1 to the cell-culture dish. Avoid incomplete removal of the cell-culture medium in order to allow full lysis activity of the lysis buffer. Continue with lysate filtration (step 3 of the standard protocol).
- **Adherent cell cultures (lysis after trypsinization):**
  Aspirate cell-culture medium and wash cells once with PBS. Aspirate PBS. Add 0.1 – 0.3 % trypsin in PBS and incubate for an appropriate time to detach the cells from the dish surface. After cell detachment, add medium, transfer cells to an appropriate tube (not supplied), and pellet by centrifugation for 5 min at 300 x *g*. Remove supernatant and continue with the addition of Lysis Buffer RA1 to the cell pellet.

- **Animal tissues:**
  It is essential for efficient RNA preparation that all the RNA contained in the sample is released from the cells by disruption and that the viscosity of the sample is reduced by homogenization.
  Thawing of undisrupted animal tissue should only be done in the presence of Buffer RA1 under simultaneous mechanical disruption, for example with a rotor-stator homogenizer or a bead mill. This ensures that the RNA is not degraded by RNases before the preparation has started.
  Commonly used techniques for disruption of animal tissues are, for example grinding with **pestle and mortar** or using **a syringe and needle** for multiple passage of the sample through the needle. However, due to the small size of samples to be processed with **NucleoSpin® RNA XS** these disruption methods are often not suitable.

2.4 Elution procedures

A high RNA concentration in the elution fraction is desirable for all typical downstream applications. In particular with regard to limited volumes of reaction mixtures, high RNA concentration can be a crucial criterion. Due to a high default elution volume, standard kits often result in weakly concentrated RNA, if only small samples are processed.

Such RNA often even requires a subsequent concentration to be suitable for the desired application.

In contrast to standard kits, **NucleoSpin® RNA XS** allows an efficient elution in a very small volume resulting in highly concentrated RNA.

Elution volumes in the range of 5 – 30 µL are recommended, the default volume is 10 µL.

2.5 Stability of isolated RNA

Eluted RNA should immediately be put and always kept on ice during work for optimal stability! Contamination with almost omnipresent RNases (general lab ware, fingerprints, dust) may be a risk for isolated RNA. For short term storage freeze at -20 °C, for long term storage freeze at -70 °C.

* Add TCEP optional before or after freezing.

3 Storage conditions and preparation of working solutions

Attention: *Buffers RA1, RA2, and MDB contain chaotropic salt and detergents. Wear gloves and goggles!*

CAUTION: Buffers RA1, RA2, and MDB contain guanidinium thiocyanate which can form highly reactive compounds when combined with bleach (sodium hypochlorite). DO NOT add bleach or acidic solutions directly to the sample-preparation waste.

- Store lyophilized **rDNase, Reducing Agent TCEP**, and **Carrier RNA** at 4 °C on arrival (stable up to 1 year).
- All other kit components should be stored at 15 – 25 °C and are stable until: see package label. Storage at lower temperatures may cause precipitation of salts.
- Check that 70 % ethanol is available as additional solution in the lab to adjust RNA binding conditions in the Buffer RA1 lysate.
- Check that 96 – 100 % ethanol is available (necessary for clean up protocol only).

Before starting with any **NucleoSpin® RNA XS** protocol prepare the following:

- **rDNase:** Add indicated volume (see following table or label on the rDNase vial) of RNase-free H_2O to the rDNase vial and incubate for 1 min at room temperature. Gently swirl the vials to completely dissolve the rDNase. Be careful not to mix rDNase vigorously as rDNase is sensitive to mechanical agitation. Dispense into aliquots and store at -20 °C. The frozen working solution is stable for 6 months. Do not freeze/thaw the aliquots more than three times.
- **Reducing Agent TCEP:** Add indicated volume of RNase-free H_2O to the TCEP vial and incubate for several minutes at room temperature. Mix the vial to completely dissolve the TCEP. Store dissolved TCEP at -20 °C.
- **Carrier RNA:** Prepare a **stock solution** before first time using: Dissolve the Carrier RNA* in 750 µL Buffer RA1 to obtain a 400 ng/µL stock solution. Prepare a **working solution** before RNA extraction: Dilute 1:100 with Buffer RA1 (e.g., 1 µL Carrier RNA stock solution + 99 µL Buffer RA1) to obtain the working solution of 4 ng/µL. Add 5 µL of this working solution (20 ng) to every lysate (protocol step 3 in section 5). Store stock solution at -20 °C; do not store working solution, prepare it freshly immediately before use.
  Note: Due to the production procedure and the small amount of Carrier RNA contained in the vial, the carrier may hardly be visible .
- **Wash Buffer RA3:** Add the indicated volume of 96 – 100 % ethanol to Wash Buffer RA3 Concentrate. Mark the label of the bottle to indicate that ethanol was added. Store Wash Buffer RA3 at 15 – 25 °C for up to one year.

* Due to the production procedure, lyophilized Carrier RNA might hardly be visible in the vial.

NucleoSpin® RNA XS			
REF	**10 preps** **740902.10**	**50 preps** **740902.50**	**250 preps** **740902.250**
Wash Buffer RA3 (Concentrate)	6 mL Add 24 mL ethanol	12 mL Add 48 mL ethanol	50 mL Add 200 mL ethanol to each bottle
rDNase, RNase-free (lyophilized)	1 vial (size A) Add 55 µL RNase-free H_2O	1 vial (size C) Add 230 µL RNase-free H_2O	2 vials (size D) Add 540 µL RNase-free H_2O to each vial
Carrier RNA	300 µg	300 µg	300 µg
	Add 750 µL Buffer RA1 to obtain concentrated stock solution. Dilute 1:100 with Buffer RA1 to obtain working solution.		
Reducing Agent TCEP	14 mg Add 100 µL RNase-free H_2O	3 × 14 mg Add 100 µL RNase-free H_2O to each vial	2 × 107 mg Add 750 µL RNase-free H_2O to each vial

4 Safety instructions

When working with the **NucleoSpin® RNA XS** kit wear suitable protective clothing (e.g., lab coat, disposable gloves, and protective goggles). For more information consult the appropriate Material Safety Data Sheets (MSDS available online at ***www.mn-net.com/msds***).

Caution: Guanidinium thiocyanate in buffer RA1 and buffer RA2 can form highly reactive compounds when combined with bleach! Thus, do not add bleach or acidic solutions directly to the sample preparation waste.

The waste generated with the **NucleoSpin® RNA XS** kit has not been tested for residual infectious material. A contamination of the liquid waste with residual infectious material is highly unlikely due to strong denaturing lysis buffer treatment but it cannot be excluded completely. Therefore, liquid waste must be considered infectious and should be handled and discarded according local safety regulations.

4.1 Disposal

Dispose hazardous, infectious or biologically contaminated materials in a safe and acceptable manner and in accordance with all local and regulatory requirements.

5 Protocols

5.1 RNA purification from cultured cells, laser captured cells, or microdissected cryosections

Before starting the preparation:

- Check if TCEP, Carrier RNA, rDNase, and Wash Buffer RA3 were prepared according to section 3.

1 Supply sample

Provide sample such as a pellet of up to 10^5 cultured cells, laser captured cells or microdissected cryosections in a microcentrifuge tube (not provided).

For appropriate sample amounts see section 2.2.

2 Lyse and homogenize cells

Add **100 µL Buffer RA1** and **2 µL TCEP** to the cell sample and vortex vigorously (2 × 5 s).

+ 100 µL RA1
+ 2 µL TCEP

If multiple samples are processed, the preparation of a master-premix is recommended (e.g., 1.1 mL Buffer RA1 and 22 µL TCEP for 10 preparations). Use 102 µL of the premix.

This procedure is usually sufficient to homogenize cultured cells, laser captured cells, or microdissected cryosections. For further comments on homogenization methods see section 2.3.

3 Add Carrier RNA

Add **5 µL Carrier RNA working solution** (20 ng) to the lysate. Mix by vortexing (2 × 5 s). Spin down briefly (approx. 1 s 1000 x *g*) to clear the lid.

+ 5 µL Carrier RNA

Mix

For preparation of Carrier RNA working solution see section 3

4 Filtrate lysate (*optional*)

Place a **NucleoSpin® Filter** (violet ring) in a Collection Tube (2 mL; supplied), apply the mixture, and centrifuge for **30 s** at **11,000 x *g***.

11,000 x *g*
30 s

This step may be skipped when working with small amounts of sample, for example less than 10^5 cells.

5 Adjust RNA binding condition

Discard the **NucleoSpin® Filter** (violet ring). Add **100 µL ethanol (70 %)** to the homogenized lysate and mix by pipetting up and down (5 times).

Alternatively, add 100 µL ethanol (70 %) to the sample in a 1.5 mL microcentrifuge tube (not provided) and mix by vortexing (2 × 5 s). Spin down briefly (approx. 1 s 1000 x g) to clear the lid. Pipette lysate up and down two times before loading the lysate.

+ 100 µL 70 % EtOH

Mix

6 Bind RNA

For each preparation, take one **NucleoSpin® RNA XS Column** (light blue ring) placed in a Collection Tube. **Load** the lysate to the column. Centrifuge for **30 s** at **11,000 x *g***.

Place the column in a new Collection Tube (2 mL).

The maximum loading capacity of NucleoSpin® RNA XS Columns is 600 µL. Repeat the procedure if larger volumes are to be processed.

Load lysate

11,000 x *g*
30 s

7 Desalt silica membrane

Add **100 µL MDB** (Membrane Desalting Buffer) and centrifuge at **11,000 x *g*** for **30 s** to dry the membrane. It is not necessary to use a fresh Collection Tube after this centrifugation step.

Salt removal will make the following rDNase digest much more effective. If the column outlet has come into contact with the flowthrough for any reason, discard the flowthrough and centrifuge again for 30 s at 11,000 x g.

+ 100 µL MDB

11,000 x *g*
30 s

8 Digest DNA

Prepare **rDNase reaction mixture** in a sterile microcentrifuge tube (not provided): for each isolation, add **3 µL** reconstituted rDNase (also see section 3) to **27 µL Reaction Buffer for rDNase**. Mix by flicking the tube.

Apply **25 µL rDNase reaction mixture** directly onto the center of the silica membrane of the column. Close the lid. Incubate at **room temperature** for **15 min**.

It is not necessary to use a new Collection Tube after the incubation step.

+ 25 µL rDNase reaction mixture

RT
15 min

9 Wash and dry silica membrane

1st wash

Add **100 µL Buffer RA2** to the NucleoSpin® RNA XS Column. Incubate for **2 min** at **RT**. Centrifuge for **30 s** at **11,000 x *g***.

Place the column into a new Collection Tube (2 mL).

Buffer RA2 will inactivate the rDNase.

+ 100 µL RA2

RT
2 min

11,000 x *g*
30 s

2nd wash

Add **400 µL Buffer RA3** to the NucleoSpin® RNA XS Column. Centrifuge for **30 s** at **11,000 x *g***. Discard flowthrough and place the column back into the Collection Tube.

+ 400 µL RA3

11,000 x *g*
30 s

Note: Make sure that residual buffer from the previous steps is washed away with Buffer RA3, especially if the lysate has been in contact with the inner rim of the column during loading of the lysate onto the column. For efficient washing of the inner rim flush it with Buffer RA3.

3rd wash

Add **200 µL Buffer RA3** to the NucleoSpin® RNA XS Column. Centrifuge for **2 min** at **11,000 x *g*** to dry the membrane. Place the column into a nuclease-free Collection Tube (1.5 mL; supplied).

If for any reason the liquid level in the Collection Tube has reached the NucleoSpin® RNA XS Column after centrifugation, discard flowthrough and centrifuge again.

+ 200 µL RA3

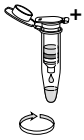
11,000 x *g*
2 min

10 Elute highly pure RNA

Elute the RNA in **10 µL H_2O** (RNase-free; supplied) and centrifuge at **11,000 x *g*** for **30 s**.

If higher RNA concentrations or higher elution volumes are desired, elution volume may be varied in the range of 5 – 30 µL.

For further details on alternative elution procedures see section 2.4.



+ 10 µL RNase-free H_2O

11,000 x *g*
30 s

5.2 RNA purification from tissue

Before starting the preparation:

- Check if TCEP, Carrier RNA, rDNase, and Wash Buffer RA3 were prepared according to section 3.

1 Supply sample

Provide tissue sample such as a biopsy in a microcentrifuge tube (not provided).

For appropriate sample amounts see section 2.2.

2 Lyse and homogenize tissue

Add **200 µL Buffer RA1** and **4 µL TCEP** to the tissue sample and vortex vigorously (2 × 5 s).

+ 200 µL RA1
+ 4 µL TCEP

Disruption with a rotor-stator homogenizer or with a shaker and steel balls are recommended methods for the homogenization of tissue samples. For further comments on homogenization methods see section 2.3.

If multiple samples are processed, the preparation of a master-premix is recommended (e.g., 2.2 mL Buffer RA1 and 44 µL TCEP for 10 preparations). Use 204 µL of the premix.

3 Add Carrier RNA

Add **5 µL Carrier RNA working solution** (20 ng) to the lysate. Mix by vortexing (2 × 5 s). Spin down briefly (approx. 1 s 1000 x *g*) to clear the lid.

+ 5 µL Carrier RNA

Mix

For preparation of Carrier RNA working solution see section 3.

4 Filtrate lysate

Reduce viscosity and clear the lysate by filtration through **NucleoSpin® Filter** (violet ring): Place the NucleoSpin® Filter (violet ring) in a Collection Tube (2 mL; provided), apply the mixture, and centrifuge for **30 s** at **11,000 x *g***.

11,000 x *g*
30 s

In case of visible pellet formation (depending on sample amount and nature), transfer supernatant without any formed pellet to a new 1.5 mL microcentrifuge tube (not included).

5 Adjust RNA binding condition

Discard the NucleoSpin® Filter (violet ring), add **200 µL ethanol (70 %)** to the homogenized lysate and mix by pipetting up and down (5 times).

Alternatively, transfer flowthrough into a new 1.5 mL microcentrifuge tube (not provided), add **200 µL ethanol** (70 %), and mix by vortexing (2 × 5 s). Spin down briefly (approx. 1 s 1000 x *g*) to clear the lid. Pipette lysate up and down two times before loading the lysate.

After addition of ethanol a stringy precipitate may become visible which will not affect the RNA isolation. Be sure to disaggregate any precipitate by mixing and load all of the precipitate on the column as described in step 6. Do not centrifuge the ethanolic lysate before loading it onto the column in order to avoid pelleting the precipitate.

+ 200 µL 70 % EtOH

Mix

6 Bind RNA

For each preparation, take one **NucleoSpin® RNA XS Column** (light blue ring) placed in a Collection Tube and load the lysate to the column. Centrifuge for **30 s** at **11,000 x *g***. Place the column in a new Collection Tube (2 mL).

The maximum loading capacity of NucleoSpin® RNA XS Columns is 600 µL. Repeat the procedure if larger volumes are to be processed.

Load lysate

11,000 x *g*
30 s

7 Desalt silica membrane

Add **100 µL MDB** (Membrane Desalting Buffer) and centrifuge at **11,000 x *g*** for **30 s** to dry the membrane. It is not necessary to use a fresh Collection Tube after this centrifugation step.

Salt removal will make the following rDNase digest much more effective. If the column outlet has come into contact with the flowthrough for any reason, discard the flowthrough and centrifuge again for 30 s at 11,000 x g.

+ 100 µL MDB

11,000 x *g*
30 s

8 Digest DNA

Prepare **rDNase reaction mixture** in a sterile microcentrifuge tube (not provided): for each isolation, add **3 µL** reconstituted rDNase (also see section 3) to **27 µL Reaction Buffer for rDNase**. Mix by flicking the tube.

Apply **25 µL rDNase reaction mixture** directly onto the center of the silica membrane of the column. Close the lid. Incubate at **room temperature** for **15 min**.

It is not necessary to use a new Collection Tube after the incubation step.

+ 25 µL rDNase reaction mixture

RT
15 min

9 Wash and dry silica membrane

+ 100 µL RA2

RT
2 min

11,000 x *g*
30 s

Add **100 µL Buffer RA2** to the NucleoSpin® RNA XS Column. Incubate for **2 min** at **RT**. Centrifuge for **30 s** at **11,000 x *g***.

Place the column into a new Collection Tube (2 mL).

Buffer RA2 will inactivate the rDNase.

+ 400 µL RA3

11,000 x *g*
30 s

Add **400 µL Buffer RA3** to the NucleoSpin® RNA XS Column. Centrifuge for **30 s** at **11,000 x *g***. Discard flowthrough and place the column back into the Collection Tube.

+ 200 µL RA3

11,000 x *g*
2 min

Add **200 µL Buffer RA3** to the NucleoSpin® RNA XS Column. Centrifuge for **2 min** at **11,000 x *g*** to dry the membrane. Place the column into a nuclease-free Collection Tube (1.5 mL; supplied).

If for any reason the liquid level in the Collection Tube has reached the NucleoSpin® RNA XS Column after centrifugation, discard flowthrough and centrifuge again.

10 Elute highly pure RNA

+ 10 µL RNase-free H_2O

11,000 x *g*
30 s

Elute the RNA in **10 µL H_2O** (RNase-free; supplied) and centrifuge at **11,000 x *g*** for **30 s**.

If higher RNA concentrations or higher elution volumes are desired, elution volume may be varied in the range of 5 –30 µL.

For further details on alternative elution procedures see section 2.4.

5.3 Clean up and concentration of RNA

Before starting the preparation:

- Check if Wash Buffer RA3 were prepared according to section 3.

1 Supply sample

Sample

Provide **up to 300 µL sample** such as prepurified RNA (e.g., phenol purified) or RNA from reaction mixtures (e.g., labelling reactions) in a microcentrifuge tube (not provided).

For appropriate sample amounts see section 2.2.

2	**Prepare lysis-binding buffer premix** For every 100 µL of sample combine **25 µL Buffer RA1** with **75 µL ethanol (96 – 100 %)** and mix. If processing multiple samples, the preparation of a master-premix (1 volume Buffer RA1 plus 3 volumes ethanol 96 – 100 %) is recommended.	**+ 25 µL RA1** **+ 75 µL EtOH (96 – 100 %) per 100 µL sample** **Mix**
3	**Add Carrier RNA** Not necessary!	
4	**Filtrate lysate** Not necessary!	
5	**Adjust RNA binding condition** Add **one volume of premix** to the sample (e.g., 100 µL premix to a 100 µL sample) and mix (2 × 5 s). If necessary, spin down briefly (approx. 1 s 1000 x *g*) to clear the lid.	 **Add 1 vol. premix to sample** **Mix**
6	**Bind RNA** For each preparation, take one **NucleoSpin® RNA XS Column** (light blue ring) placed in a Collection Tube and **load the lysate** to the column. Centrifuge for **30 s** at **11,000 x *g***. For samples > 300 µL, load in two steps. Place the column in a new Collection Tube (2 mL). *For high demanding applications, the recovery rate can be increased as follows: Centrifuge 30 s at 2,000 x g prior to centrifugation for 30 s at 11,000 x g.*	 **Load lysate** **11,000 x *g* 30 s**
7	**Desalt silica membrane** Not necessary!	
8	**Digest DNA** Not necessary!	

9 **Wash and dry silica membrane**

1st wash

Add **400 µL Buffer RA3** to the NucleoSpin® RNA XS Column. Centrifuge for **30 s** at **11,000 x *g***. Discard flowthrough and place the column back into the Collection Tube.

+ 400 µL RA3

11,000 x *g*
30 s

2nd wash

Add **200 µL Buffer RA3** to the NucleoSpin® RNA XS Column. Centrifuge for **2 min** at **11,000 x *g*** to dry the membrane. Place the column into a nuclease-free Collection Tube (1.5 mL; supplied).

If for any reason the liquid level in the Collection Tube has reached the NucleoSpin® RNA XS Column after centrifugation, discard flowthrough and centrifuge again.

+ 200 µL RA3

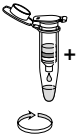
11,000 x *g*
2 min

10 **Elute highly pure RNA**

Elute the RNA in **10 µL H_2O** (RNase-free; supplied) and centrifuge at **11,000 x g** for **30 s**.

If higher RNA concentrations or higher elution volumes are desired, elution volume may be varied in the range of 5 – 30 µL.

For further details on alternative elution procedures see section 2.4.



+ 10 µL RNase-free H_2O

11,000 x *g*
30 s

5.4 rDNase digestion in the eluate

The on column rDNase digestion in the standard protocol is very efficient and thus results in minimal residual DNA. This DNA will not be detectable in most downstream applications. However, removal of DNA to a completely undetectable level is challenging and the efficiency of an on column DNA digestion is sometimes not sufficient for downstream applications requiring lowest residual content of DNA.

A typical example for such a demanding application is an RT-PCR reaction in which the primer molecules do not differentiate between cDNA (derived from RNA) and contaminating genomic DNA. Especially, if

- high copy number targets are analyzed (e.g., multi gene family, mitochondrial, plastidal or plasmid targets (from transfections))
- the target gene is of a very low expression level
- the amplicon is relatively small (< 200 bp).

DNA digestion in solution can efficiently destroy contaminating DNA. However, stringent RNase control and subsequent repurification of the RNA (in order to remove buffer, salts, DNase and digested DNA) are usually required.

The high quality, recombinant, RNase-free DNase (rDNase) in the **NucleoSpin® RNA XS** kits facilitates such a digestion in solution in order to remove even traces of contaminating DNA.

A **Digest DNA (Reaction setup)**

Prepare enzyme-buffer premix: Add **1 µL rDNase** to **10 µL Reaction Buffer for rDNase.**

Add 1/10 volume of enzyme-buffer premix to the eluted RNA (e.g., to 10 µL RNA add 1 µL of the premix comprising buffer and enzyme).

B **Incubate sample**

Incubate for **10 min** at **37 °C**.

C **Repurify RNA**

Repurify RNA with a suitable RNA cleanup procedure, for example following section 2.4, by ethanol precipitation or with the NucleoSpin® RNA Clean-up XS kit (see ordering information).

Ethanol precipitation, exemplary

Add 0.1 volume of 3 M sodium acetate, pH 5.2 and 2.5 volumes of 96 – 100 % ethanol to one volume of sample. Mix thoroughly.

Incubate several minutes to several hours at -20 °C or 4 °C, respectively.

Note: Choose long incubation times if the sample contains low RNA concentration. Short incubation times are sufficient if the sample contains high RNA concentration.

Centrifuge for 10 min at max. speed.

Wash RNA pellet with 70 % ethanol.

Dry RNA pellet and resuspend RNA in RNase-free H_2O.

6 Appendix

6.1 Troubleshooting

Problem	Possible cause and suggestions
RNA is degraded/ no RNA obtained	*RNase contamination* • Create an RNase free working environment. Wear gloves during all steps of the procedure. Change gloves frequently. Use of sterile, disposable polypropylene tubes is recommended. Keep tubes closed whenever possible during the preparation. Glassware should be oven-baked for at least 2 hours at 250 °C before use.
Poor RNA quality or yield	*Reagents not applied or restored properly* • Reagents not properly restored. Add the indicated volume of RNase-free H_2O to rDNase vial and 96 % ethanol to Buffer RA3 Concentrate and mix. Reconstitute and store lyophilized rDNase according to instructions given in section 3. • Sample and reagents have not been mixed completely. Always vortex vigorously after each reagent has been added. • No ethanol has been added after lysis. Binding of RNA to the silica membrane is only effective in the presence of ethanol. *Kit storage* • Reconstitute and store lyophilized rDNase according to instructions given in section 3. • Store other kit components at room temperature. Storage at low temperatures may cause salt precipitation. • Keep bottles tightly closed in order to prevent evaporation or contamination. *Ionic strength and pH influence A_{260} absorption as well as ratio A_{260}/A_{280}* • For adsorption measurement, use 5 mM Tris pH 8.5 as diluent. Please see also: - Manchester, K L. 1995. Value of A_{260}/A_{280} ratios for measurement of purity of nucleic acids. Biotechniques 19, 208 – 209. - Wilfinger, W W, Mackey, K and Chomczyski, P. 1997. Effect of pH and ionic strength on the spectrophotometric assessment of nucleic acid purity. Biotechniques 22, 474 – 481.

Problem	Possible cause and suggestions
Poor RNA quality or yield (continued)	*Sample material* • Sample material not stored properly. Whenever possible, use fresh material. If this is not possible, store samples in NucleoProtect® RNA stabilization solution or flash freeze the samples in liquid N_2. Store unprotected samples at -70 °C. Never allow unprotected samples to thaw before addition of Buffer RA1. Perform disruption of samples immediately after addition of Lysis Buffer RA1. • Insufficient disruption and / or homogenization of starting material. Ensure thorough sample disruption and use NucleoSpin® Filters for easy homogenization of disrupted starting material.
Low A_{260}/A_{230} ratio	*Carry-over of guanidinium thiocyanate* • Carefully load the lysate to the NucleoSpin® RNA XS Column and try to avoid a contamination of the upper part of the column and the column lid. • Make sure that residual Wash Buffer RA2 is washed away with Wash Buffer RA3. This may be done by applying Buffer RA3 to the inner rim of the column.
Clogged NucleoSpin® Column/ Poor RNA quality or yield	*Sample material* • Too much starting material used. Overloading may lead to decreased overall yield. Reduce amount of sample material or use larger volume of Buffer RA1. • Insufficient disruption and/or homogenization of starting material. Ensure thorough sample disruption and use NucleoSpin® Filters for easy homogenization of disrupted starting material.
Contamination of RNA with genomic DNA	*rDNase not active* • Reconstitute and store lyophilized rDNase according to instructions given in section 3. *DNase solution not properly applied* • Pipette rDNase solution directly onto the center of the silica membrane and close the lid. *Too much cell material used* • Reduce quantity of cells or tissue used.

Problem	Possible cause and suggestions
Contamination of RNA with genomic DNA *(continued)*	*DNA detection system too sensitive* • The amount of DNA contamination is effectively reduced during the on column digestion with rDNase. Anyhow, it can not be guaranteed that the purified RNA is 100 % free of DNA, therefore in very sensitive applications it might still be possible to detect DNA. The probability of DNA detection with PCR increases with: - the number of DNA copies per preparation: single copy target < plastidial/mitochondrial target < plasmid transfected into cells - decreasing of PCR amplicon size. • Use larger PCR targets (e.g., > 500 bp) or intron spanning primers if possible. • **Use support protocol 5.4 for subsequent rDNase digestion in solution.**
Suboptimal performance of RNA in downstream experiments	*Carry-over of ethanol or salt* • Do not let the flowthrough touch the column outlet after the second Buffer RA3 wash. Be sure to centrifuge at the corresponding speed for the respective time in order to remove ethanolic Buffer RA3 completely. • Check if Buffer RA3 has been equilibrated to room temperature before use. Washing at lower temperatures lowers efficiency of salt removal by Buffer RA3. • Depending on the robustness of the used RT-PCR system, RT-PCR might be inhibited if complete eluates are used as template for RT-PCR. Use less eluate as template. *Store isolated RNA properly* • Eluted RNA should always be kept on ice for optimal stability since trace contaminations of omnipresent RNases (general lab ware, fingerprints, dust) will degrade the isolated RNA. For short term storage freeze at -20 °C, for long term storage freeze at -70 °C.

Problem	Possible cause and suggestions
Discrepancy between A_{260} quantification values and PCR quantification values	*Silica abrasion from the membrane* • Due to the typically low RNA content in very small samples and the resulting low total amount of isolated RNA, an RNA quantification via A_{260} absorption measurement is often hampered due to the low sensitivity of the absorption measurement. When performing absorption measurements close to the detection limit of the photometer, the measurement may be influenced by minor amounts of silica abrasion. In order to prevent incorrect A_{260}-quantification of small RNA amounts centrifuge the eluate for 30 s at > 11.000 x *g* and take an aliquot for measurement without disturbing any sediment. Alternatively, use a silica abrasion insensitive RNA quantification method (e.g., RiboGreen fluorescent dye).
Unexpected A_{260}/A_{280} ratio	*Measurement not in the range of photometer detection limit* • In order to obtain a significant A_{260}/A_{280} ratio it is necessary that the initially measured A_{260} and A_{280} values are significantly above the detection limit of the photometer used. An A_{280} value close to the background noise of the photometer will cause unexpected A_{260}/A_{280} ratios.

6.2 Ordering information

Product	REF	Pack of
NucleoSpin® RNA XS	740902.10/.50/.250	10/50/250
NucleoSpin® RNA Clean-up XS	740903.10/.50/.250	10/50/250
NucleoSpin® totalRNA FFPE XS	740969.10/.50/.250	10/50/250
NucleoSpin® RNA	740955.10/.50/.250	10/50/250
NucleoZOL	740404.200	200 mL
NucleoSpin® RNA Blood	740200.10/.50	10/50
NucleoSpin® totalRNA FFPE	740982.10/.50/.250	10/50/250
NucleoSpin® RNA Midi	740962.20	20
NucleoSpin® RNA/Protein	740933.10/.50/.250	10/50/250
NucleoSpin® TriPrep	740966.10/.50/.250	10/50/250
NucleoSpin® RNA Clean-up	740948.10/.50/.250	10/50/250
NucleoSpin® RNA/DNA Buffer Set	740944	Suitable for 100 preps
Buffer RA1	740961 740961.500	60 mL 500 mL
rDNase Set	740963	1 set
Reducing Agent TCEP	740395.107	107 mg
NucleoSpin® Filters	740606	50
Collection Tubes (2 mL)	740600	1000
NucleoProtect® RNA	740400.50/.250/.500	50/250/500 mL

Visit ***www.mn-net.com*** for more detailed product information.

6.3 References

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Imbeaud S, Graudens E, Boulanger V, Barlet X, Zaborski P, Eveno E, Mueller O, Schroeder A, Auffray C.: Towards standardization of RNA quality assessment using user-independent classifiers of microcapillary electrophoresis traces. Nucleic Acids Res. 2005 Mar 30;33(6):e56.

Miller CL, Diglisic S, Leister F, Webster M, Yolken RH.: Evaluating RNA status for RT-PCR in extracts of postmortem human brain tissue. Biotechniques. 2004 Apr; 36(4):628 – 33.

Schoor O, Weinschenk T, Hennenlotter J, Corvin S, Stenzl A, Rammensee HG, Stevanovic S.: Moderate degradation does not preclude microarray analysis of small amounts of RNA. Biotechniques. 2003 Dec; 35(6):1192 – 6, 1198 – 201.

6.4 Product use restriction / warranty

All MACHEREY-NAGEL products are designed for their intended use only. They are not intended to be used for any other purpose. The description of the intended use of the products can be found in the original MACHEREY-NAGEL product leaflets. Before using our products, please observe the instructions for use and the safety instructions from the respective Material Safety Data Sheet of the product.

This MACHEREY-NAGEL product is carrying documentation stating specifications and other technical information. MACHEREY-NAGEL warrants to meet the stated specifications. The provided warranty is limited to the data specifications and descriptions as given in the original MACHEREY-NAGEL literature. No other statements or representations, written or oral, by MACHEREY-NAGEL's employees, agents or representatives, except written statements signed by a duly authorized officer of MACHEREY-NAGEL are authorized. They should not be relied upon by the costumer and are not a part of a contract of sale or of this warranty.

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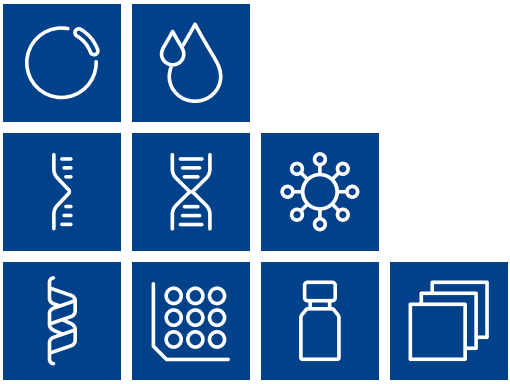

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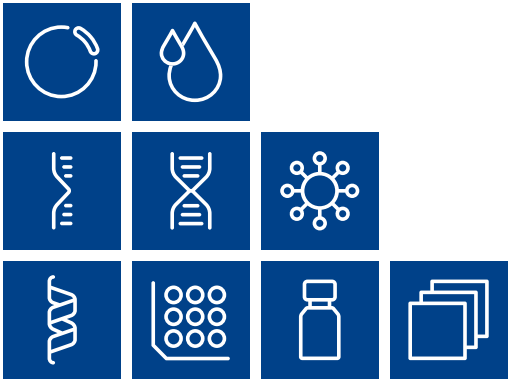

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