

NucleoSpin® Tissue

1 Kit contents

NucleoSpin® Tissue		
REF	12 preps 740952.12c	240 preps 740952.240c
Lysis Buffer T1	5 mL	50 mL
Lysis Buffer B3	15 mL	75 mL
Wash Buffer BW	30 mL	2 × 125 mL
Wash Buffer B5 (Concentrate)	6 mL (add 24 mL ethanol before first use)	100 mL (add 100 mL ethanol to each bottle before first use)
Elution Buffer BE	30 mL	125 mL
Proteinase K (lyophilized)	10 mg (add 430 µL Proteinase Buffer PB for reconstitution)	2 × 75 mg (add 3.35 mL Proteinase Buffer PB for reconstitution to each vial)
Proteinase Buffer PB	1.8 mL	8 mL
NucleoSpin® Tissue Columns (light green rings)	12	240
User manual	1	1

2 How to use the kit

Refer to the protocol information on the following pages for technical details on using the kit. For further questions or detailed instructions, particularly regarding the use of the kit with specific robotic instruments, please contact MACHEREY-NAGEL at support@mn-net.com.

NucleoSpin® kits on QIAcube®

MN is not recommending to use this kit on specific robots. The use of NucleoSpin® kits on the QIAcube® is solely at your own discretion. MACHEREY-NAGEL is not responsible for loss of warranty claims or other consequences.

3 General information

Application:	DNA
Kit:	NucleoSpin® Tissue (REF 740952.240c) instead of: DNeasy® Blood & Tissue Kit
Sample material:	Tissue and rodent tails
Protocol name:	total DNA from animal blood and tissues and from cells, yeast, bacteria, or viruses DNA_DNeasyBloodAndTissue_TissueAndRodentTails_Standard_ V1
Short protocol name:	Standard

4 Using the kit

1. Lyse up to 25 mg animal tissue in 180 µL Buffer T1 and 20 µL Proteinase K solution. When using 0.4–0.6 cm rodent tail clippings use one clipping for rats or two for mouse. Incubate 6h or overnight at 56 °C until the tissue is completely digested.

If necessary, centrifuge after lysis to pellet any remaining unlysed debris and transfer the clear lysate to a new tube.

For details refer to NucleoSpin® Tissue user manual.

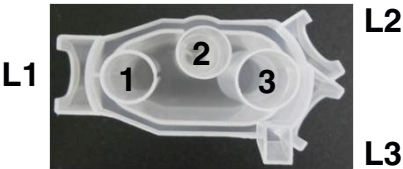
2. Fill the designated buffer bottles with the buffers according to the buffer table on page 3.
3. Transfer 200 µL of each pre-lysed sample to a 2 mL Sample Tube RB.
4. Place sample tubes into sample rack.
5. Insert disposable Filter-Tips 1000 µL and 1000 µL wide-bore.
6. General equipment setup is shown on page 3.

5 Additional materials

Refer to the QIAcube® protocol sheet for required consumables (e.g., sample tubes, collection tubes, instrument accessories, disposable tips, etc.) and software requirements.

6 Rotor adapter

Position	Labware	Lid position
1	NucleoSpin® Tissue Column	L1
2	–	–
3	1.5 mL collection tube*	L3



* Sarstedt, Micro tube 1.5 mL Safety Cap

7 Buffers (Reagent Bottle Rack)

Position	MN Reagent	Replaced QIAGEN® Reagent
1	–	–
2	Buffer B3	Buffer B3
3	96 – 100 % ethanol	96 – 100 % ethanol
4	Buffer BW	Buffer AW1
5	Buffer B5	Buffer AW2
6	Buffer BE	Buffer AE

Comment: Use a pipet to transfer the required volume of Lysis Buffer B3 to the reagent bottle. Avoid generating large air bubbles or foam as this may lead to a liquid detection error during the load check.

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