

## QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:



[www.mn-net.com/sds](http://www.mn-net.com/sds)



We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact our Technical Support.

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## User manuals

**NucleoMag® DNA/RNA Water**      744220 / .1 / .4



Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned product are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols, can all be found on the product website and accessed easily via the QR code.

### QR-Code product website



[qr.mn-net.com/qr/\(241\)744220](https://qr.mn-net.com/qr/(241)744220)

Use the following QR code or the link below for direct access to the user manual.

### QR-Code user manual
















[qr.mn-net.com/qr/\(IFU\)744220](https://qr.mn-net.com/qr/(IFU)744220)

It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This leaflet does not replace the full manual!

Protocol at a glance - Isolation of DNA/RNA from water samples  
 (25 mm filter membrane)

200 $\mu$ L sample		
<b>1 Filter water sample</b>		Filter 100 – 1000 mL of a water sample through a filter membrane (25 mm)
<b>2 Lyse sample</b>		Insert filter membrane into a MN Bead Tube Type A 500 $\mu$ L MWA1 Agitate on a swing mill or similar device 0.5 – 15 min 11,000 x g, 30 s
<b>3 Bind DNA/RNA to NucleoMag<sup>®</sup>R B-Beads</b>	 	Up to 450 $\mu$ L lysate 25 $\mu$ L NucleoMag <sup>®</sup> B-Beads 475 $\mu$ L MWA2  Mix by shaking for 5 min at RT <i>(Optional: Mix by pipetting up and down)</i>  Remove supernatant after 5 min separation
<b>4 Wash with MWA3</b>	 	Remove Square-well Block from NucleoMag <sup>®</sup> SEP 850 $\mu$ L MWA3  Resuspend: Shake 2 min at RT <i>(Optional: Mix by pipetting up and down)</i>  Remove supernatant after 2 min separation
<b>5 Wash with MWA3</b>	 	Remove Square-well Block from NucleoMag <sup>®</sup> SEP 850 $\mu$ L MWA3  Resuspend: Shake 2 min at RT <i>(Optional: Mix by pipetting up and down)</i>  Remove supernatant after 2 min separation
<b>6 Wash with MWA4</b>	 	Remove Square-well Block on NucleoMag <sup>®</sup> SEP 850 $\mu$ L MWA4  Resuspend: Shake 2 min at RT <i>(Optional: Mix by pipetting up and down)</i>  Remove supernatant after 2 min separation
<b>7 Dry the beads</b>		10 – 15 min at RT
<b>8 Elute DNA</b>	 	Remove Square-well Block from NucleoMag <sup>®</sup> SEP 50-100 $\mu$ L RNase-free H <sub>2</sub> O <i>(Optional: Elute at 56 °C)</i>  Shake 5 min at RT <i>(Optional: Mix by pipetting up and down)</i>  Separate 2 min and transfer DNA into elution plate

