

User manuals

NucleoBond® Xtra Midi	740410/.10/.50/.100
NucleoBond® Xtra Midi Plus	740412/.10/.50
NucleoBond® Xtra Maxi	740414/.10/.50/.100
NucleoBond® Xtra Maxi Plus	740416/.10/.50



Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned products are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols, can all be found on the product website and accessed easily via the QR code.

QR-Code product website



[qr.mn-net.com/qr/\(241\)740410](https://qr.mn-net.com/qr/(241)740410)

Use the following QR code or the link below for direct access to the user manual.

QR-Code user manual



[qr.mn-net.com/qr/\(IFU\)740410](https://qr.mn-net.com/qr/(IFU)740410)

It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This leaflet does not replace the full manual!

Protocol at a glance (Rev.17)
Plasmid DNA purification (NucleoBond® Xtra Midi / Maxi)

Midi		
1 – 3	Cultivation and harvest	
4 – 5	Cell lysis <i>(Important: Check Buffer LYS for precipitated SDS)</i>	High-copy / low-copy 8 mL / 16 mL 8 mL / 16 mL Buffer RES. Buffer LYS RT, 5 min
6	Equilibration of the column and filter	12 mL Buffer EQU
7	Neutralization	8 mL / 16 mL Buffer NEU Mix thoroughly until colorless
8	Clarification and loading of the lysate	
9	1 st Wash	5 mL Buffer EQU
10	Filter removal	Discard NucleoBond® Xtra Column Filter
11	2 nd Wash	8 mL Buffer WASH
12	Elution	5 mL Buffer ELU
13	Precipitation	NucleoBond® Xtra MidiNucleoBond®
		3.5 mL Isopropanol Vortex 4,5 – 15,000 x g 4 °C, 30 min
14	Washing and drying	2 mL 70 % ethanol 4,5 – 15,000 x g RT, 5 min 10 – 15 min
15	Reconstitution	Appropriate volume of TE buffer

Maxi		
4,500 – 6,000 x g 4 °C, 15 min		
	High-copy / low-copy	
	12 mL / 24 mL 12 mL / 24 mL	Buffer RES Buffer LYS
	RT, 5 min	
	25 mL Buffer EQU	
	12 mL / 24 mL	Buffer NEU
	Mix thoroughly until colorless	
Invert the tube 3 times		
Load lysate on NucleoBond® Xtra Column Filter		
	15 mL Buffer EQU	!
	Discard NucleoBond® Xtra Column Filter	
!	25 mL Buffer WASH	!
	15 mL Buffer ELU	
Xtra Midi Plus	NucleoBond® Xtra Maxi	NucleoBond® Xtra Maxi Plus
15 mL Isopropanol Vortex 2 min NucleoBond® Finalizer	10.5 mL Isopropanol Vortex 4,5 – 15,000 x g 4 °C, 30 min	10.5 mL Isopropanol Vortex RT, 2 min Load NucleoBond® Finalizer Large
4 mL 70 % ethanol	4 mL 70 % ethanol 4,5 – 15,000 x g RT, 5 min	4 mL 70 % ethanol
until dry	15 – 30 min	≥ 6 x air until dry
800 µL Buffer TRIS	Appropriate volume of TE buffer	400 – 1000 µL Buffer TRIS

QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:



www.mn-net.com/sds



We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact our Technical Support.

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