

User manuals

NucleoBond® Xtra Midi EF	740420/.10/.50
NucleoBond® Xtra Midi Plus EF	740422/.10/.50
NucleoBond® Xtra Maxi EF	740424/.10/.50
NucleoBond® Xtra Maxi Plus EF	740426/.10/.50



Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned products are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols, can all be found on the product website and accessed easily via the QR code.

QR-Code product website



[qr.mn-net.com/qr/\(241\)740420](https://qr.mn-net.com/qr/(241)740420)

Use the following QR code or the link below for direct access to the user manual.

QR-Code user manual



[qr.mn-net.com/qr/\(IFU\)740420](https://qr.mn-net.com/qr/(IFU)740420)

It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This leaflet does not replace the full manual!

Protocol at a glance (Rev.12)
Endotoxin-free plasmid DNA purification
(NucleoBond® Xtra Midi EF/Maxi EF)

Midi			
1 – 3	Cell cultivation and harvest		
4 – 5	Cell lysis <i>(Important: Check Buffer LYS for precipitated SDS)</i>	High-copy / low-copy 8 mL / 16 mL Buffer RES-EF 8 mL / 16 mL Buffer LYS-EF RT, 5 min	
6	Equilibration of the column and filter	15 mL Buffer EQU-EF	
7	Neutralization	8 mL / 16 mL Buffer NEU-EF Mix thoroughly until colorless Incubate 5 min on ice	
8	Clarification and loading of the lysate		
9	1 st Wash	5 mL Buffer FIL-EF	
10	Filter removal	Discard NucleoBond® Xtra Column Filter	
11	2 nd Wash	35 mL Buffer ENDO-EF	
12	3 rd Wash	15 mL Buffer WASH-EF	
13	Elution	5 mL Buffer ELU-EF	
14	Precipitation	NucleoBond® Xtra Midi EF	NucleoBond®
		3.5 mL Isopropanol Vortex 4,5 – 15,000 x g 4 °C, 30 min	3 Iso V R Load N Final
15	Washing and drying	2 mL 70 % EtOH 4,5 – 15,000 x g RT, 5 min 10 – 15 min	70 ≥ 6 x
16	Reconstitution	Appropriate volume of TE-EF or H ₂ O-EF	200 TE-EF

Maxi		
4,500 – 6,000 x g 4 °C, 15 min		
	High-copy / low-copy 12 mL / 24 mL Buffer RES-EF 12 mL / 24 mL Buffer LYS-EF RT, 5 min	
	35 mL Buffer EQU-EF	
	12 mL / 24 mL Buffer NEU-EF Mix thoroughly until colorless Incubate 5 min on ice	
Invert the tube 3 times Load lysate on NucleoBond® Xtra Column Filter		
	10 mL Buffer FIL-EF	
	Discard NucleoBond® Xtra Column Filter	
	90 mL Buffer ENDO-EF	
	45 mL Buffer WASH-EF	
	15 mL Buffer ELU-EF	
NucleoBond® Xtra Midi Plus EF	NucleoBond® Xtra Maxi EF	NucleoBond® Xtra Maxi Plus EF
10.5 mL Isopropanol Vortex RT, 2 min Load NucleoBond® Finalizer Large	10.5 mL Isopropanol Vortex 4,5 – 15,000 x g 4 °C, 30 min	10.5 mL Isopropanol Vortex RT, 2 min Load NucleoBond® Finalizer Large
5 mL 70 % EtOH Air until dry	5 mL 70 % EtOH 4,5 – 15,000 x g RT, 5 min 15 – 30 min	5 mL 70 % EtOH ≥ 6 x air until dry
400 – 800 µL TE-EF or H ₂ O-EF	Appropriate volume of TE-EF or H ₂ O-EF	400 – 1000 µL TE-EF or H ₂ O-EF

QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:



www.mn-net.com/sds



We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact our Technical Support.

Contact MN

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