

## User manuals

### NucleoSpin® DNA FFPE XS 740980.10 / .50 / .250



Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned product are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols, can all be found on the product website and accessed easily via the QR code.

#### QR-Code product website



[qr.mn-net.com/qr/\(241\)740980](https://qr.mn-net.com/qr/(241)740980)

Use the following QR code or the link below for direct access to the user manual.

#### QR-Code user manual



[qr.mn-net.com/qr/\(IFU\)740980](https://qr.mn-net.com/qr/(IFU)740980)










It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This leaflet does not replace the full manual!














## Protocol at a glance (Rev. 04)

## DNA Isolation from FFPE Samples

DNA isolation with Paraffin Dissolver		
<b>Sample preparation</b>		For appropriate sample quantity see section 2.4.
<b>1 Deparaffinize sample</b>		400 $\mu$ L Paraffin Dissolver 60 $^{\circ}$ C, 3 min Mix hot sample
		Let sample cool down
<b>2 Lyse sample</b>		100 $\mu$ L FL Mix vigorously 11,000 x g, 1 min 10 $\mu$ L Proteinase K Mix lower phase RT, 3 hours or overnight
<b>3 Decrosslink</b>		100 $\mu$ L D-Link Mix gently 11,000 x g, 30 s 90 $^{\circ}$ C, 30 min
<b>4 Adjust binding conditions</b>		200 $\mu$ L ~ 98 % ethanol Mix 1,000 x g, 1 s
<b>5 Bind DNA</b>		Load aqueous (lower) phase 2,000 x g, 30 s
<b>6 Wash and dry silica membrane</b>		
<b>1<sup>st</sup> wash</b>		400 $\mu$ L B5 11,000 x g, 30 s
<b>2<sup>nd</sup> wash</b>		400 $\mu$ L B5 11,000 x g, 2 min
<b>7 Elute DNA</b>		20 $\mu$ L BE 11,000 x g, 30 s
<b>8 Optional: Remove residual ethanol</b>		90 $^{\circ}$ C, 8 min

## DNA isolation with xylene

For appropriate sample quantity see section 2.4.

	1 mL xylene RT, 2 min Mix
	11,000 x g, 2 min Discard supernatant 1 mL ~ 98 % ethanol Mix
	11,000 x g, 2 min Discard supernatant
	Dry at 60 °C, 3–10 min
	100 µL FL – 10 µL Proteinase K Mix RT, 3 hours or overnight
	100 µL D-Link Mix gently – 90 °C, 30 min
	200 µL ~ 98 % ethanol Mix –
 	Load lysate 2,000 x g, 30 s
	
	400 µL B5 11,000 x g, 30 s
	400 µL B5 11,000 x g, 2 min
 	20 µL BE 11,000 x g, 30 s
	90 °C, 8 min

## QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:



[www.mn-net.com/sds](http://www.mn-net.com/sds)



We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact our Technical Support.

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