

User manuals

NucleoMag® HMW DNA

744160.1

Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned product are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols, can all be found on the product website and accessed easily via the QR code.

QR-Code product website



[qr.mn-net.com/qr/\(241\)744160](https://qr.mn-net.com/qr/(241)744160)

Use the following QR code or the link below for direct access to the user manual.

QR-Code user manual



[qr.mn-net.com/qr/\(IFU\)744160](https://qr.mn-net.com/qr/(IFU)744160)

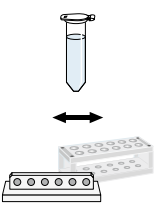
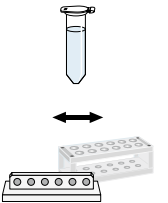
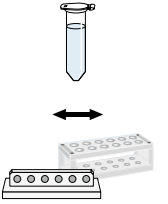
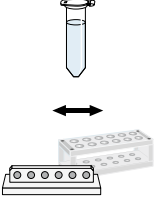
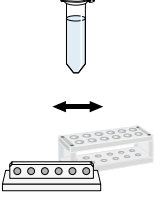
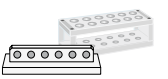
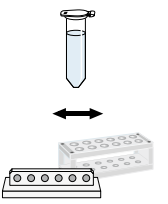
It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This leaflet does not replace the full manual!

Protocol at a glance

Isolation of high molecular weight DNA

1 Prepare and lyse samples		25 μ L Proteinase K 600 μ L HM1 Mix or invert 56 °C, 30 - 150 min
2 Precipitate debris		11,000 x g, 1 min, 5 min Transfer up to 500 μ L to a fresh 2 mL tube
3 Digest RNA		20 μ L RNase Mix or invert 15 min, RT
4 Bind DNA to NucleoMag® B-Beads		25 μ L NucleoMag® B-Beads 400 μ L HM2 Mix by shaking for 5 min at 1000 rpm (Optional: Mix by pipetting up and down using wide-bore tips) Remove supernatant after 5 min separation
5 Wash with HM3		Remove tubes from NucleoMag® SEP Mini 900 μ L HM3 Mix by shaking for 2 min, 1000 rpm (Optional: Mix by pipetting up and down using wide-bore tips) Remove supernatant after 2 min separation
6 Wash with HM4		Remove tubes from NucleoMag® SEP Mini 900 μ L HM4 Mix by shaking for 2 min, 1000 rpm (Optional: Mix by pipetting up and down using wide-bore tips) Remove supernatant after 2 min separation
7 1st Wash with 70 % EtOH		Remove tubes from NucleoMag® SEP Mini 900 μ L 70 % EtOH Mix by shaking for 2 min, 1000 rpm (Optional: Mix by pipetting up and down using wide-bore tips) Remove supernatant after 2 min separation
8 2nd wash with 70 % EtOH		Remove tubes from NucleoMag® SEP Mini 900 μ L 70 % EtOH Mix by shaking for 2 min, 1000 rpm (Optional: Mix by pipetting up and down using wide-bore tips) Remove supernatant after 2 min separation
9 Rinse the beads		Leave tubes on NucleoMag® SEP Mini 600 μ L HM5 Incubate for 30–60 s <i>Note: Do not resuspend the beads in Buffer HM5!</i> Remove supernatant
10 Elute DNA		Remove tubes from NucleoMag® SEP Mini 100 μ L HM6 Shake 5 min at RT 1000 rpm (Optional: Mix by pipetting up and down) using wide-bore tips Separate 2 min and transfer DNA into elution plate/tubes

QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:



www.mn-net.com/sds



We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact our Technical Support.

Contact MN

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