

User manuals

NucleoBond® PC 500 EF	740550
NucleoBond® PC 2000 EF	740549
NucleoBond® PC 10000 EF	740548
NucleoBond® PC Prep 100	740594



Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned products are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols, can all be found on the product website and accessed easily via the QR code.

QR-Code product website



qr.mn-net.com/qr/(241)740548

Use the following QR code or the link below for direct access to the user manual.

QR-Code user manual



qr.mn-net.com/qr/(IFU)740548

It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

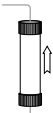
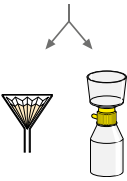
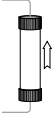
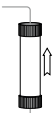
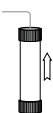
We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This is a short protocol and does not replace the full manual!

Protocol at a glance (Rev. 19) – Endotoxin-free plasmid DNA purification (I)

	Maxi (AX 500)	Mega (AX 2000)
1 Cultivation and harvest of bacterial cells	4,500–6,000 x g 15 min at 4 °C	4,500–6,000 x g 15 min at 4 °C
2 Cells lysis		
Buffer S1-EF	12 mL	40 mL
Buffer S2-EF	12 mL < 5 min at RT	40 mL < 5 min at RT
Buffer S3-EF	12 mL 5 min at 0 °C	40 mL /
		Mix thoroughly – invert 10–15 times
3 Equilibration of the column	Buffer N ₂ -EF 5 mL	Buffer N ₂ -EF 25 mL
4 Clarification of the lysate	Folded Filter 20 min	Bottle Top Filter Type 1 5 min
5 Binding	Load cleared lysate onto the column	Load cleared lysate onto the column
6 Washing	Buffer N3-EF 2 x 24 mL Buffer N4-EF 2 x 12 mL	Buffer N3-EF 1 x 60 mL 2 x 40 mL Buffer N4-EF 60 mL
7 Elution	Buffer N5-EF 15 mL	Buffer N5-EF 25 mL
8 Precipitation	Isopropanol 11 mL 4,5–15,000 x g 30 min at 12 °C	Isopropanol 18 mL 4,5–15,000 x g 30 min at 12 °C
9 Washing and drying	70 % ethanol 5 mL 4,5–15,000 x g 10 min at RT 10–20 min	70 % ethanol 7 mL 4,5–15,000 x g 10 min at RT 30–60 min
10 Reconstitution	Appropriate volume of TE-EF or H ₂ O-EF	Appropriate volume of TE-EF or H ₂ O-EF

Maxi, Mega, Giga, Preparative scale)

Giga (AX 10000)		Preparative scale (AX Prep 100)	
4,500–6,000 x <i>g</i> 15 min at 4 °C		4,500–6,000 x <i>g</i> 15 min at 4 °C	
120 mL 120 mL < 5 min at RT 120 mL /		1000 mL 1000 mL < 3 min at RT 1000 mL 25 min at 0 °C	
Buffer N ₂ -EF 100 mL		Buffer N ₂ -EF 900 mL	
Bottle Top Filter Type 2 5 min		1 st Sieving Fabric 2 nd Folded Filters	
Load cleared lysate onto the column		Load cleared lysate onto the column	
Buffer N3-EF 4 x 150 mL Buffer N4-EF 3 x 130 mL		Buffer N3-EF 900 mL Buffer N4-EF 1900 mL	
Buffer N5-EF 100 mL		Buffer N5-EF 470–600 mL	
Isopropanol 70 mL 4,5–15,000 x <i>g</i> 30 min at 12 °C	 	Isopropanol 0.7 vol 4,5–15,000 x <i>g</i> 60 min at 12 °C	 
70 % ethanol 10 mL 4,5–15,000 x <i>g</i> 10 min at RT 30–60 min	 	70 % ethanol 25 mL 4,5–15,000 x <i>g</i> 20 min at RT 60 min	 
Appropriate volume of TE-EF or H ₂ O-EF		Appropriate volume of TE-EF or H ₂ O-EF	

QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:



www.mn-net.com/sds



We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact our Technical Support.

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