

User manuals

NucleoMag® 384 Plant

744402 / .1 / .4



Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned product are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols, can all be found on the product website and accessed easily via the QR code.

QR-Code product website



[qr.mn-net.com/qr/\(241\)744402](https://qr.mn-net.com/qr/(241)744402)

Use the following QR code or the link below for direct access to the user manual.

QR-Code user manual



[qr.mn-net.com/qr/\(IFU\)744402](https://qr.mn-net.com/qr/(IFU)744402)

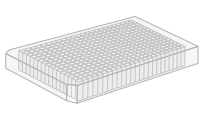
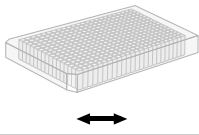
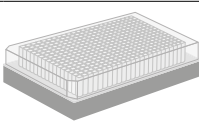
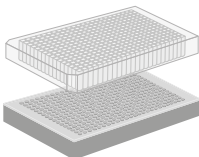
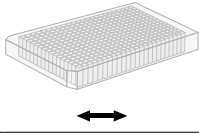
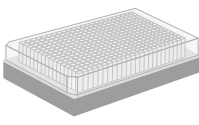
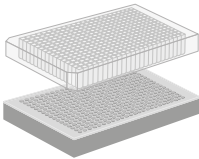
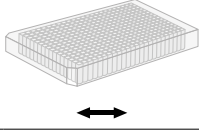
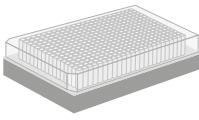
It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This leaflet does not replace the full manual!

Protocol at a glance

Protocol for the isolation of genomic DNA from plant samples in a 384-well format

1 Homogenize and lyse plant sample material (~30 mg)	Add 200 μ L MC1 and 10 μ L RNase A Mix 56 °C, 60 min	
2 Clear lysates by centrifugation, transfer 50 μL of cleared lysate for further processing	Centrifuge at 5,600 x g, 3 min Transfer 50 μ L cleared lysate	
3 Bind DNA to NucleoMag[®] V-Beads	Add 4 μ L NucleoMag [®] V-Beads and 50 μ L MC2 <i>(Note: Resuspend the NucleoMag[®] V-Beads before removing them from the storage bottle)</i>	
	Mix by shaking for 5 min at RT <i>(Optional: Mix by pipetting up and down)</i>	
	Remove supernatant after 3 min separation	
4 Wash with MC3	Remove separation plate from magnetic separator Add 80 μ L MC3	
	Resuspend: Shake 5 min at RT <i>(Optional: Mix by pipetting up and down)</i>	
	Remove supernatant after 2 min separation	
5 Wash with MC4	Remove separation plate from magnetic separator Add 80 μ L MC4	
	Resuspend: Shake 5 min at RT <i>(Optional: Mix by pipetting up and down)</i>	
	Remove supernatant after 2 min separation	

6 Wash with 80 % ethanol	Remove separation plate from magnetic separator Add 80 μ L 80 % ethanol	
	Resuspend: Shake 5 min at RT (Optional: Mix by pipetting up and down)	
	Remove supernatant after 2 min separation	
7 Wash with MC5	Leave Square-well Block on the magnetic separator! Add 80 μL MC5 Incubate for 45 – 60 s Note: Do not resuspend the beads in Buffer MC5!	
	Remove supernatant	
8 Elute DNA	Remove separation plate from magnetic separator Add 60 μ L MC6 (Elution volume possible from 50 – 200 μ L; Optional: Elute at 55 °C)	
	Shake 10 min at RT (Optional: Mix by pipetting up and down)	
	Separate 2 min and transfer DNA (supernatant) into elution plate / tubes	

QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:



www.mn-net.com/sds



We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact our Technical Support.

Contact MN

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