

## User manuals

**NucleoSpin® 8 Plant II** 740669 / .5

**NucleoSpin® 8 Plant II Core Kit** 740467.4



Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned product are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols can be found on the product website and accessed easily via the QR code.

### QR-Code product website



[qr.mn-net.com/qr/\(241\)740669](https://qr.mn-net.com/qr/(241)740669)

Use the following QR code or the link below for direct access to the user manual.

### QR-Code user manual



[qr.mn-net.com/qr/\(IFU\)740669](https://qr.mn-net.com/qr/(IFU)740669)

It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This is a short protocol and does not replace the full manual!

## Protocol at a glance

## NucleoSpin® 8 Plant II – centrifuge processing

Please check the user manual if your centrifuge and setup meets the requirements for centrifuge processing of 8-well strips.

<b>1 Homogenize samples</b>	Up to 100 mg wet or 20 mg lyophilized plant tissue 5,600 – 6,000 x g, 2 min
<b>2 a Cell lysis using Buffer PL1</b>	500 µL PL1 10 µL RNase A Mix 65 °C, 30 min Proceed with step 3
<b>2 b Cell lysis using Buffer PL2 and PL3</b>	400 µL PL2 10 µL RNase A Mix 65 °C, 30 min 100 µL PL3 Mix and incubate on ice for 5 min Proceed with step 3
<b>3 Clear lysate by centrifugation</b>	5,600 – 6,000 x g, 20 min
<b>4 Adjust binding conditions</b>	Mix 450 µL PC with 400 µL cleared lysate
<b>5 Transfer lysate to NucleoSpin® Plant II Binding Strips</b>	
<b>6 Bind DNA to silica membrane of the NucleoSpin® Plant II Binding Strips</b>	5,600 – 6,000 x g, 2 min
<b>7 Wash and dry silica membrane</b>	400 µL PW1 5,600 – 6,000 x g, 2 min 700 µL PW2 5,600 – 6,000 x g, 2 min 700 µL PW2 5,600 – 6,000 x g, 10 min
<b>8 Elute DNA</b>	100 µL PE (70 °C) (incubate 2 min) 5,600 – 6,000 x g, 2 min Repeat once

## Protocol at a glance

### NucleoSpin® 8 Plant II – vacuum processing

<b>1 Homogenize samples</b>	Up to 100 mg wet or 20 mg lyophilized plant tissue 5,600 – 6,000 x g, 2 min
<b>2 a Cell lysis using Buffer PL1</b>	500 µL PL1 10 µL RNase A Mix 65 °C, 30 min Proceed with step 3
<b>2 b Cell lysis using Buffer PL2 and PL3</b>	400 µL PL2 Mix 65 °C, 30 min 100 µL PL3 Mix and incubate on ice for 5 min Proceed with step 3
<b>3 Clear lysate by centrifugation</b>	5,000 – 6,000 x g 20 min
<b>4 Adjust binding conditions</b>	Mix 450 µL PC with 400 µL cleared lysate
<b>5 Transfer lysate to NucleoSpin® Plant II Binding Strips</b>	
<b>6 Bind DNA to silica membrane of the NucleoSpin® Plant II Binding Strips</b>	-0.2 to -0.4 bar* (2 min)
<b>7 Wash and dry silica membrane</b>	400 µL PW1 700 µL PW2 700 µL PW2 – 0.4 bar* (1 min each step) Remove MN Wash Plate Dry silica membrane (10 min, maximum vacuum)
<b>8 Elute DNA</b>	100 µL PE (incubate 2 min) -0.4 bar* (2 min) Repeat once

## QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:



[www.mn-net.com/sds](http://www.mn-net.com/sds)



We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact our Technical Support.

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