

User manuals

**NucleoSpin® DNA
Forensic**

740840 / .10 / .50 / .250



Dear valued customer,

Thank you for choosing MACHEREY-NAGEL for your application. We have attached a short protocol for your review.

To obtain the best results we recommend to follow the detailed protocol available online, especially when you are a first time user of this kit.

All important links to the above mentioned product are listed in this leaflet.

Important information regarding product components, specifications, safety instructions, and processing protocols, can all be found on the product website and accessed easily via the QR code.

QR-Code product website



[qr.mn-net.com/qr/\(241\)740840](https://qr.mn-net.com/qr/(241)740840)

Use the following QR code or the link below for direct access to the user manual.

QR-Code user manual



[qr.mn-net.com/qr/\(IFU\)740840](https://qr.mn-net.com/qr/(IFU)740840)
















It is strongly recommended to read the detailed protocol section of the user manual if using the kit for the first time. However, experienced users may refer to the protocol at a glance. The protocol at a glance is designed to be used only as a supplemental tool for quick reference while performing the purification procedure.

We are constantly improving our products and we reserve the right to make changes or additions to protocols. Please check for updated revisions for previously downloaded manuals.

This leaflet does not replace the full manual!












Protocol at a glance

Protocol for the isolation of genomic DNA from forensic samples

1 Lyse sample		Add 20 μ L Liquid Proteinase K, 5 μ L TCEP solution (14 mg/mL) and 450 μ L Buffer FOL Mix 56 °C, 1 h
2 Adjust DNA binding conditions		Add 580 μ L FOB Vortex
3 Bind DNA	 	Load 600 μ L lysate 1 min at 11,000 x <i>g</i> .
		Load remaining lysate 1 min at 11,000 x <i>g</i> .
4 Wash with FOW1	 	Add 400 μ L FOW1 1 min at 11,000 x <i>g</i> .
5 Wash with FOW2		Add 400 μ L FOW2
1 st wash		1 min at 11,000 x <i>g</i> .
6 Wash with FOW2		Add 400 μ L FOW2
2 nd wash		1 min at 11,000 x <i>g</i> .
7 Dry silica membrane	 	2 min at 11,000 x <i>g</i> .
8 Elute DNA	 	Add 50 – 100 μ L Buffer FOE. 1 min RT 1 min at 11,000 x <i>g</i> .

Protocol at a glance

Protocol for the isolation of genomic DNA from bone or teeth samples

Sample preparation		Mill up to 150 mg of bone or teeth to a fine powder. Transfer up to 150 mg of bone powder into a 2 mL microcentrifuge tube.
1 Pre-lyse samples		Add 200 μ L phosphate buffer (0.5 M EDTA/0.25 M PO ₄ ³⁻ , pH 8), 700 μ L Buffer T1 and 20 μ L Proteinase K. Mix 56 °C shaking overnight. Afterwards 48 h at 2–8 °C on a shaking incubator.
2 Lyse sample		Mix, Add 800 μ L Buffer B3 Mix 10 min, 70 °C Mix 10 min 5,000 x g. Transfer 800 μ L of the supernatant to a new 2 mL microcentrifuge tube.
3 Adjust binding conditions		Add 1160 μ L Binding Buffer FOB Mix
4 Bind DNA to NucleoSpin® DNA Forensic Colum		Load 600 μ L lysate 1 min 11,000 x g Discard the flowthrough and place the column back in the Collection Tube.
5 Repeat binding step		Load 600 μ L 1 min 11,000 x g Discard the flowthrough and place the column back in the Collection Tube.
6 Load the remaining lysate to the column		1 min at 11,000 x g Discard the flowthrough and place the column in a new Collection Tube (provided)
7 Wash with FOW1		Add 400 μ L Buffer FOW1 to the column and centrifuge for 1 min at 11,000 x g. Discard the flowthrough and place the column back into the Collection Tube.
8 Wash with FOW2 (1st)		Add 400 μ L Buffer FOW2 1 min 11,000 x g
9 Wash with FOW2 (2nd)		Add 400 μ L Buffer FOW2 1 min 11,000 x g
10 Dry silica membrane		2 min 11,000 x g
11 Elute DN		Add 100 μ L Buffer FOE 1 min at room temperature 1 min 11,000 x g

QR-Code Safety Data Sheet

Please find a digital version of the safety data sheets by following the link below:



www.mn-net.com/sds



We strongly recommend to carefully read the detailed protocol section of the product's user manual. If you have any questions about the protocol or product, please contact our Technical Support.

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