

MACHEREY-NAGEL

User manual



Purification of His-tag proteins

- Protino® 96 Ni-NTA

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1 Components

1.1 Contents and storage

| Protino® 96 Ni-NTA | | |
|---|----------|------------|
| REF | 745425.1 | 745425.4 |
| Protino® Ni-NTA Agarose (sufficient for 96 preps using an agarose bed volume of 50 µL for the purification of up to 2 mg polyhistidine tagged protein) | 5.5 mL | 4 x 5.5 mL |
| Protino® Purification Plate (proprietary leak-free purification plate for retaining the chromatographic resins) | 1 | 4 |
| MN Wash Plate (one plate is used to minimize cross-contamination, the other plate is used as an adaptor to place the Protino® Purification Plate on a plate shaker) | 2 | 8 |
| User manual | 1 | 1 |

Shipping and storage

! The product is shipped at ambient temperature.

■ Upon receipt, Protino® Ni-NTA Agarose suspension should be **stored at 2–8 °C** and is stable up to 1 year. Do not freeze.

All other components can be stored at ambient temperature.

1.2 Additional material to be supplied by user

- Plate shaker
- Vacuum manifold system OR centrifuge
- Plate(s) for collecting eluate fraction(s) (Square-well Block, see ordering information)
- For the purification under native conditions, prepare the buffers according section 4.1.
- For the purification under denaturing conditions, prepare buffers according section 5.1.

2 Product description

2.1 The basic principle

Overview

Protino® Ni-NTA products enable convenient high throughput purification of recombinant polyhistidine-tagged proteins by immobilized metal ion affinity chromatography (IMAC). Proteins from any expression system can be purified under native or denaturing conditions using Protino® Ni-NTA Agarose.

Individual amounts of the IMAC matrix can be loaded into the wells of the Protino® Purification Plate. However, we recommend to use 50 µL bed volume per well. Due to the special designed filter-frit material, the Protino® Purification Plate is absolutely leak-free in operation. Therefore, it is possible to perform the entire purification process (equilibration, batch-binding, washing, elution) directly in the wells of the purification plate. Buffer passes through filter-frits only by vacuum or centrifugation. The Protino® Purification Plate enables high well-to-well reproducibility. In addition, the long drip directors of the plate minimize cross-contamination between the wells.

The uniquely designed Protino® Purification Plate is also ideal for parallel screening experiments to optimize chromatographic conditions including the sample-to-resin ratio, sample incubation time, wash and elution conditions, buffer additives, etc.

Protino® Ni-NTA technology

Binding of protein is based on the interaction between the polyhistidine tag of the recombinant protein and immobilized Ni²⁺ ions. Protino® Ni-NTA Agarose consists of the chelating ligand nitrilotriacetic acid (NTA) immobilized on 6 % crosslinked agarose beads that are suitable for batch binding, gravity flow, and FPLC™ columns. The resin is precharged with Ni²⁺ ions and therefore ready to use. Protino® Ni-NTA Agarose uses NTA which represents the most commonly used chelating ligand in IMAC. NTA is a tetradentate chelator which occupies four out of the six binding sites in the coordination sphere of the Ni²⁺ ion. The remaining two coordination sites are usually occupied by water molecules and can be exchanged with histidine residues of the recombinant protein (Figure 1). This formation of coordination sites has turned out to be optimal for purification of polyhistidine-tagged proteins. The result is two available binding sites in the coordination sphere of the Ni²⁺ ion that enable tight but reversible selective protein interactions. Chelation of Ni²⁺ ions by NTA through four coordination positions minimizes metal leaching during purification and increases specificity for polyhistidine-tagged proteins.

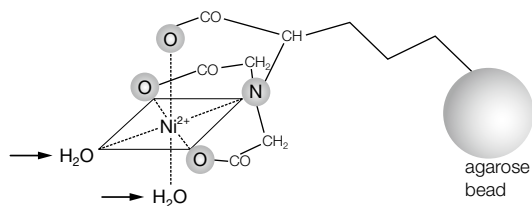


Figure 1: Protino® Ni-NTA Agarose – structure of NTA in complex with Ni²⁺

2.2 Specifications

Table 1: Specifications Protino® 96 Ni-NTA

| | |
|------------------------------------|---|
| Purification plate material | Polypropylene |
| Filter-frits | According to special designed filter-frit material, the filter plates are absolutely leak-free. Buffer passes through filter-frits only when the water breakthrough pressure is reached by vacuum or centrifugation |
| Filter diameter | 8.3 mm |
| Number of wells | 96 |
| Volume capacity of well | 1.4 mL |
| Recommended sample volume | Up to 750 µL (at 1,100 rpm on an eppendorf Thermomixer®) |
| Matrix | Protino® Ni-NTA Agarose, crosslinked 6% beaded agarose |
| Form | 50% aqueous suspension containing 30 vol% ethanol, precharged with Ni ²⁺ |
| Ligand | Nitrilotriacetic acid (NTA) |
| Bead size | 45–165 µm |
| Bed volume | Variable |
| Recommended bed volume | 50 µL |
| Binding capacity* | Up to 2 mg per well when using 50 µL settled agarose |

* Binding capacity will vary for each polyhistidine-tagged protein and strongly depends on chromatographic conditions.

Table 1: Specifications Protino® 96 Ni-NTA

| | |
|---|---|
| Recommended vacuum | -0.6 bar (depends on sample properties) |
| Recommended centrifugation speed | 3,000 x <i>g</i> (depends on sample properties) |

2.3 General information

Binding capacity

The binding capacity of Protino® Ni-NTA Agarose strongly depends on the characteristics of the polyhistidine-tagged protein including amino acid composition, molecular weight, 3-D structure, oligomerization properties, etc. Protein yield also depends on the total amount and concentration of the target protein in the sample which in turn directly correlate with the expression level and the cell density of the expression culture. Furthermore, chromatographic conditions may effect protein binding, such as sample-to-resin ratio, sample incubation time, wash and elution conditions, buffer additives, etc.

For optimal results, we recommend to use 50 µL of settled Protino® Ni-NTA Agarose per well. A capacity of up to mg per well was determined for the monomeric green fluorescent protein (6 x His-GFPuv, ~32 kDa) expressed in *E. coli*.

Incubation time

Especially during sample incubation, sufficient agitation is necessary to disperse the agarose beads. We recommend to use 50 µL of settled Protino® Ni-NTA Agarose per well, an agitation speed of 1,100 rpm (eppendorf Thermomixer®) and a sample volume of up to 750 µL.

As a starting point, we recommend to shake the suspension for 15–30 min (see Figure 2). However, as the purification plate is absolutely leak-free, longer sample incubation times can be used to achieve maximum yield (see Figure 2). If shorter incubation periods are required (e.g., according to protein stability), incubation can be reduced (e.g., to 5 min), but then yield may decrease (see Figure 2).

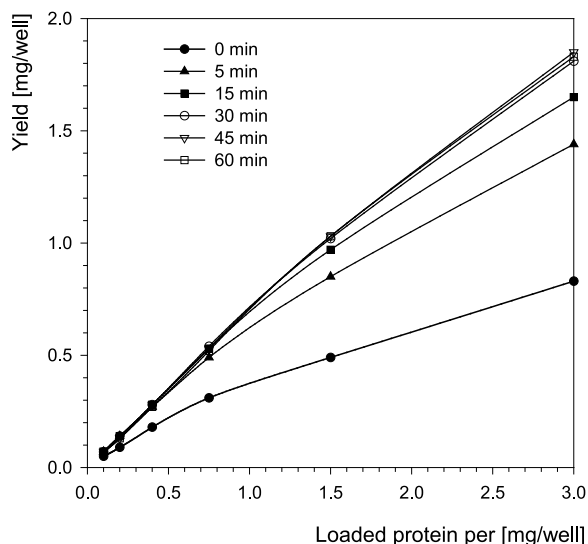


Figure 2: Impact of sample incubation time on protein yield

Protino® Ni-NTA Agarose (50 μ L bed volume) was loaded with increasing amounts of His-tagged Green Fluorescent Protein (6 x His GFPuv) in a total volume of 750 μ L of NPI-10. After washing with 1.5 mL of NPI-20, the target protein was eluted with 2 x 200 μ L of NPI-250. Yields of 6 x His-GFPuv are plotted versus the amount of loaded protein. When wells are loaded with up to 0.8 mg, shaking for 5 min already leads to acceptable yields. However, if larger protein amounts are loaded and maximum yield is required, we recommend to shake the plate for 15 to 30 min.

Solubility of the recombinant protein

Protein yield is also dependent on solubility of the recombinant protein. If proteins are expressed in *E. coli*, ideally the target proteins remain soluble in the cytoplasm. However, proteins that are highly expressed tend to accumulate in insoluble aggregates forming inclusion bodies.

For solubilization of inclusion bodies, buffers containing large amounts of denaturants are used.

This manual includes instructions for isolation of soluble proteins (purification under native conditions, see section 4) as well as insoluble proteins from inclusion bodies (purification under denaturing conditions, see section 5).

Improving purity

Sometimes optimization of purification procedures is necessary to increase purity.

Usually lysis/equilibration buffers contain 10 mM and the wash buffer 20 mM of imidazole, to suppress binding of contaminating proteins. To improve specificity, increase imidazole concentration.

In addition, for more stringent binding and washing conditions, the pH may be reduced from pH 8 closer to pH 7 (e.g., pH 7.4) in all buffers.

Additives

Avoid high concentration of additives that interact with nickel ions and thus reduce capacity (e.g., chelating agents (EDTA) or reducing agents (DTT, mercaptoethanol)), see compatibility of reagents (section 2.4).

Cross-contamination

The MN Wash Plate prevents the bottom of the Protino® Purification Plate to be contaminated by sample or by buffers. Any spray will be drained away from the purification plate to waste. The risk for cross-contamination is highly reduced. The MN Wash Plate is discarded after the washing steps.

2.4 Compatibility of reagents

| Reagent | Effect | Comments |
|-------------------|--|---|
| Sodium phosphate | Used in buffers in order to buffer the solutions at pH 8 | 50 mM is recommended; the pH of any buffer should be adjusted to 8, although in some cases a pH between 7 and 8 can be used |
| Tris, HEPES, MOPS | Coordinates with Ni ²⁺ ions, causing a decrease in capacity | Up to 100 mM may be used, sodium phosphate buffer is recommended |
| Sodium Chloride | Prevents ionic interactions and therefore unspecific binding | Up to 2 M can be used, at least 0.3 M should be used |
| Imidazole | Binds to immobilized Ni ²⁺ ions and competes with the polyhistidine-tagged proteins | Is used at low concentration to reduce non specific binding (20 mM) and to elute the target protein (> 100 mM) |
| Urea | Solubilizes protein | Use 8 M for purification under denaturing conditions |

| | | |
|--|---|--|
| GuHCl | Solubilizes protein | Up to 6 M can be used |
| β -mercaptoethanol | Prevents formation of disulfide bonds; can reduce Ni^{2+} ions at higher concentrations | Up to 20 mM in samples has been used successfully in some cases |
| DTT, DTE | Can reduce Ni^{2+} ions at higher concentrations | Up to 10 mM in samples has been used successfully in some cases |
| Glutathione reduced | Can reduce Ni^{2+} ions at higher concentrations | Up to 30 mM in samples has been used successfully in some cases |
| Glycerol | Prevents hydrophobic interactions between proteins | Up to 50 % can be used |
| EDTA | Coordinates with Ni^{2+} ions, causing a decrease in capacity at higher concentrations | Not recommended, but up to 1 mM in samples has been used successfully in some cases |
| Ethanol | Prevents hydrophobic interactions between proteins | Up to 20 % can be used; ethanol may precipitate proteins, causing low flow rates and column clogging |
| SDS | Interacts with Ni^{2+} ions, causing a decrease in capacity | Not recommended, but up to 0.3 % in samples has been used successfully in some cases |
| Nonionic detergents: Triton, Tween, etc. | Removes background proteins | Up to 2 % can be used |


3 Safety instructions

The following components of the **Protino® 96 Ni-NTA** products contain hazardous contents.

Wear gloves and goggles and follow the safety instructions given in this section.

GHS classification

Only harmful features need not be labeled with H and P phrases until 125 mL or 125 g. *Mindergefährliche Eigenschaften müssen bis 125 mL oder 125 g nicht mit H- und P-Sätzen gekennzeichnet werden.*

| Component | Hazard contents | GHS symbol | Hazard phrases | Precaution phrases |
|----------------------------|--|---|----------------|--------------------|
| <i>Inhalt</i> | <i>Gefahrstoff</i> | <i>GHS-Symbol</i> | <i>H-Sätze</i> | <i>P-Sätze</i> |
| Protino® Ni-NTA Agarose | Ethanol (diluted) 5–20 % + resin suspension with Ni in nickel(II) complexes 0,04–0,38 % <i>Ethanol (verdünnt) 5–20 % + Polymersuspension mit Ni(II) in Nickelkomplexen 0,04–0,38 %</i> CAS 64-17-5d, MN0050 |  WARNING WARNUNG | 317 | 261, 280 |

Hazard phrases

H 317 May cause an allergic skin reaction.
Kann allergische Hautreaktionen verursachen.

Precaution phrases

P 261 Avoid breathing dust/fume/gas/mist/vapours/spray.
Einatmen von Staub/Rauch/Gas/Nebel/Dampf/Aerosol vermeiden.

P 280 Wear protective gloves/protective clothing/eye protection/face protection.
Schutzhandschuhe/Schutzkleidung/Augenschutz/Gesichtsschutz tragen.

For further information, please see Material Safety Data Sheets (www.mn-net.com).
Weiterführende Informationen finden Sie in den Sicherheitsdatenblättern (www.mn-net.com).



The symbol shown on labels refers to further safety information in this section.
Das auf Etiketten dargestellte Symbol weist auf weitere Sicherheitsinformationen dieses Kapitels hin.

4 Purification of polyhistidine-tagged proteins under native conditions

4.1 Preparation of buffers for purification under native conditions

Buffer volumes needed for 96 preps using 50 μ L of Protino[®] Ni-NTA Agarose per well.

| Step | Buffer | Volume / well | Exact volume / plate | Recommended volume / plate |
|---------------|---------|---------------|----------------------|----------------------------|
| Equilibration | NPI-10 | 1 x 0.5 mL | 48 mL | 60 mL |
| Washing | NPI-20 | 3 x 0.5 mL | 144 mL | 150 mL |
| Elution | NPI-250 | 2 x 0.2 mL | 38.4 mL | 50 mL |

Please note that additional NPI-10 buffer may be needed for the preparation of protein extracts.

NPI-10/lysis & equilibration buffer (1 liter):

| | | | | |
|--------|----------------------------------|---------|---|-------------------|
| 50 mM | NaH ₂ PO ₄ | 7.80 g | NaH ₂ PO ₄ • 2 H ₂ O | Mr = 156.01 g/mol |
| 300 mM | NaCl | 17.54 g | NaCl | Mr = 58.44 g/mol |
| 10 mM | imidazole | 0.68 g | imidazole | Mr = 68.08 g/mol |

Adjust pH to 8.0 using NaOH

NPI-20/wash buffer (1 liter):

| | | | | |
|--------|----------------------------------|---------|---|-------------------|
| 50 mM | NaH ₂ PO ₄ | 7.80 g | NaH ₂ PO ₄ • 2 H ₂ O | Mr = 156.01 g/mol |
| 300 mM | NaCl | 17.54 g | NaCl | Mr = 58.44 g/mol |
| 20 mM | imidazole | 1.36 g | imidazole | Mr = 68.08 g/mol |

Adjust pH to 8.0 using NaOH

NPI-250/elution buffer (1 liter):

| | | | | |
|--------|----------------------------------|---------|---|-------------------|
| 50 mM | NaH ₂ PO ₄ | 7.80 g | NaH ₂ PO ₄ • 2 H ₂ O | Mr = 156.01 g/mol |
| 300 mM | NaCl | 17.54 g | NaCl | Mr = 58.44 g/mol |
| 250 mM | imidazole | 17.00 g | imidazole | Mr = 68.08 g/mol |

Adjust pH to 8.0 using NaOH

4.2 Preparation of cleared *E. coli* lysates under native conditions

1 Cultivate and harvest cells

- *E. coli* cells may be cultivated in 96-well plates (1 mL culture volume), 24-well plates (5 mL culture volume), or any other appropriate cultivation vessel, if larger culture volumes are required.
 - Harvest cells by centrifugation at 2,000 x *g* for 15 min at 4 °C.
 - Store cell pellets at -20 °C or -70 °C for at least 1 h.
-

2 Prepare cell extracts

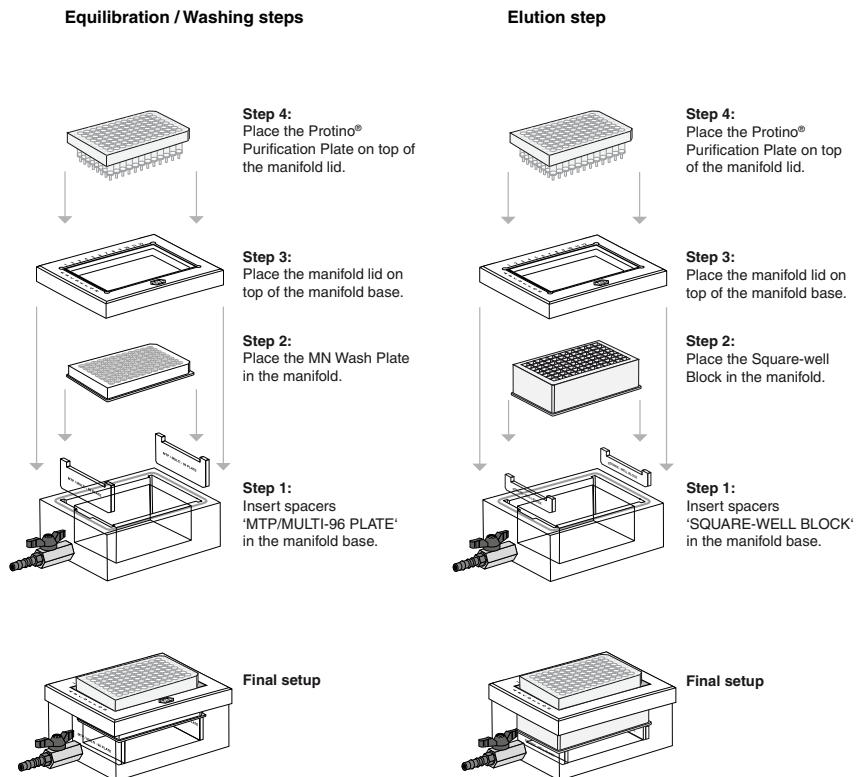
Use standard procedures for the preparation of cell extracts, such as lysozyme treatment, sonication, or detergent treatment. Note that optimal sample preparation steps have to be determined empirically depending on the characteristics of the polyhistidine-tagged protein and host organism.

For preparation of cell extracts from up to 5 mL *E. coli* expression culture, we recommend the following protocol as a starting point for further optimization:

- Thaw cell pellets at room temperature.
 - Resuspend each pellet in 1 mL of 1 x NPI-10 Buffer containing 0.2 mg/mL lysozyme.
 - Incubate at room temperature for 30 min in a shaker. If the lysate is still viscous, add 15 U of Benzonase® per well, mix, and incubate at room temperature for 30 min. Benzonase® reduces lysate viscosity by rapidly hydrolysing DNA and RNA.
 - Centrifuge the crude lysate at 5,000 x *g* for 30 min at 4 °C to remove cellular debris. If the supernatant is not clear, centrifuge a second time to avoid clogging of the Protino® Purification Plate with insoluble material.
 - Store supernatant on ice.
-

4.3 Protocol for purification under native conditions using vacuum manifold

Setting up the NucleoVac 96 Vacuum Manifold:



1 Set up the NucleoVac 96 Vacuum Manifold for the preparation of the Protino® Purification Plate

- Insert spacers labeled 'MTP/MULTI-96 PLATE' notched side up into the grooves located on the short sides of the manifold.
- Insert waste container into manifold base.
- Insert the MN Wash Plate on the spacers (labeled 'MTP/MULTI-96 PLATE') inside the manifold base (here the MN Wash Plate is used to avoid cross-contaminations).
- Close the manifold base with the manifold lid.

2 Prepare Protino® Purification Plate

- Resuspend Protino® Ni-NTA Agarose by mixing thoroughly to achieve a homogeneous suspension.
- Pour the entire contents of one bottle (11 mL of suspension) into a appropriate tray.
- Add 44 mL of deionized water to get a final volume of 55 mL .
- Gently agitate the tray to achieve a homogeneous suspension.

Note: Proper agitation is necessary to make sure the agarose beads are evenly mixed. If the suspension is not mixed well, agarose beads settle to the bottom of the tray and lead to inconsistent filling and finally to poor well-to-well reproducibility!

Use a tray with a flat bottom!

Keep mixing the agarose suspension until all wells of the plate are filled!

The suspension is considered homogeneous if it appears uniform to the eye. No clouds or settled agarose beads should be visible!

- Dispense 500 µL of the suspension into each well of the Protino® Purification Plate (500 µL of 10 % suspension corresponds to 50 µL bed volume).

Note: If you do not use all wells of the plate for purification, seal the top of the empty wells with a foil.

- Place the Protino® Purification Plate on top of the manifold base.
- Apply vacuum of approximately -0.6 bar* for 1 min. If necessary, press down the plate slightly until flow through starts.
- When the diluted storage solution of the agarose has passed the plate, release the vacuum.

*Reduction of atmospheric pressure

3 Equilibration

- Equilibrate Protino® Ni-NTA Agarose by adding 500 µL of Lysis/ Equilibration Buffer (NPI-10) to each well.
- Apply vacuum of approximately -0.6 bar* for 1 min.
- Allow the buffer to pass the wells.
- Apply vacuum of approximately -0.8 bar* for a few seconds to remove any residual fluid from the long drip directors.
- Release the vacuum.

4 Batch binding

- Add up to 750 µL of clarified *E. coli* lysate to the preequilibrated wells.
- Put a MN Wash Plate upside down into the plate shaker (here the MN Wash Plate is used as an adaptor to place the Protino® Purification Plate on a plate shaker).
- Place the Protino® Purification Plate on top of the inverted MN Wash Plate (the drips of the Protino® Purification Plate fit right into the holes of the MN Wash Plate).

Note: For robotic applications or more convenient handling, MN provides a special frame to fix the Protino® Purification Plate on plate shakers (see ordering information).

- Depending on the dimension of the shaker, fix the plates properly.
- Shake the plate at 20 °C for 15 min to 30 min.

Note: Adjust the shaking speed so that the agarose remains in suspension and that no liquid comes out of the wells (e.g., 1,100 rpm for an eppendorf Thermomixer®).

5 Lysate removal

- Place the Protino® Purification Plate on top of the manifold base.
- Apply vacuum of approximately -0.6 bar* until all wells have drained. If necessary, press down the plate slightly until flow through starts.
- *Note: The vacuum may have to be adjusted for optimal results.*
- Apply vacuum of approximately -0.8 bar* for a few seconds to remove any residual fluid from the long drip directors.
- When the *E. coli* lysate has passed the plate, release the vacuum.

*Reduction of atmospheric pressure

6 Washing

- Wash Protino® Ni-NTA Agarose by adding 500 µL of Wash Buffer (NPI-20) to each well.
- Apply vacuum of approximately -0.6 bar* for 1 min.
- Allow the buffer to pass the wells.
- Apply vacuum of approximately -0.8 bar* for a few seconds to remove any residual fluid from the long drip directors.
- Release the vacuum.
- Repeat the washing step twice (total wash 3 x 500 µL of NPI-20).
- Remove Protino® Purification Plate from the vacuum manifold.

7 Set up the NucleoVac 96 Vacuum Manifold for Elution

- Remove manifold lid, MN Wash Plate, and waste container from the vacuum manifold.
 - Insert spacers labeled 'SQUARE-WELL BLOCK' notched side up into the grooves located on the short sides of the manifold.
 - Insert a Square-well Block into manifold base.
 - Close the manifold base with the manifold lid.
-

*Reduction of atmospheric pressure

8 Elution

- Add 200 µL of Elution Buffer (NPI-250) to each well of the Protino® Purification Plate.
- Place the Protino® Purification Plate on a shaker by using the inverted MN Wash Plate from step 4 (batch binding).
- Shake the plate at 20 °C and 1,100 rpm for 1 min (Eppendorf Thermomixer®).
- Place the Protino® Purification Plate on top of the manifold base.
- Elute the polyhistidine tagged proteins by applying vacuum of approximately -0.6 bar* for 1 min.
- Apply vacuum of approximately -0.8 bar* for a few seconds to remove any residual fluid from the long drip directors.
- Release the vacuum.
- Repeat the elution step once or until all target protein is removed.
- You may collect the eluates either in the same Square-well Block or in fractions by changing the block between each elution step.
- Store eluted protein on ice.

Note: Prior further analysis, mix the eluate thoroughly.

*Reduction of atmospheric pressure

4.4 Procedure for purification under native conditions by centrifugation

- Follow the standard protocol as described in section 4.2. The vacuum steps are substituted by centrifugation of the Protino® Purification Plate at **3,000 x g** for **2 min** at RT.
 - During all centrifugation steps, the Protino® Purification Plate should be placed on a Square-well Block (see ordering information, section 6.2) to collect the waste. Omit the MN Wash Plate.
 - During the elution step, the Protino® Purification Plate is placed either on top of a Rack of MN Tube Strips (see ordering information, section 6.2) or on a Square-well Block.
-

5 Purification of polyhistidine-tagged proteins under denaturing conditions

Purification of polyhistidine-tagged proteins under denaturing conditions is similar to purification under native conditions except that the sample and the buffers contain a denaturant (8 M urea or 6 M guanidine hydrochloride).

For the preparation of cell extracts and the equilibration steps, we recommend to use 6 M guanidine hydrochloride, as it is the stronger denaturant. For the washing and elution step, we recommend to switch to 8 M urea, because then eluates can be applied directly to SDS-PAGE analysis.

5.1 Preparation of buffers for purification under denaturing conditions

Buffer volumes needed for 96 preps using 50 µL of Protino® Ni-NTA Agarose:

| Step | Buffer | Volume/well | Exact volume/plate | Recommended volume/plate |
|---------------|---|-------------|--------------------|--------------------------|
| Equilibration | Denaturing Lysis/ Equilibration Buffer | 1 x 0.5 mL | 48 mL | 60 mL |
| Washing | Denaturing Wash Buffer | 3 x 0.5 mL | 144 mL | 150 mL |
| Elution | Denaturing Elution Buffer | 2 x 0.2 mL | 38.4 mL | 50 mL |

Please note that additional Denaturing Lysis/Equilibration Buffer may be needed for the preparation of protein extracts.

Denaturing Lysis/Equilibration Buffer (1 liter):

| | | | | |
|--------|----------------------------------|---------|---|-------------------|
| 0.1 M | NaH ₂ PO ₄ | 15.6 g | NaH ₂ PO ₄ • 2 H ₂ O | Mr = 156.01 g/mol |
| 0.01 M | Tris Base | 1.2 g | Tris Base | Mr = 121.14 g/mol |
| 6 M | guanidine hydrochloride | 573.2 g | guanidine hydrochloride | Mr = 95.53 g/mol |

Adjust pH to 8.0 using NaOH

Denaturing Wash Buffer (1 liter):

| | | | | |
|--------|----------------------------------|---------|---|-------------------|
| 0.1 M | NaH ₂ PO ₄ | 15.6 g | NaH ₂ PO ₄ • 2 H ₂ O | Mr = 156.01 g/mol |
| 0.01 M | Tris Base | 1.2 g | Tris Base | Mr = 121.14 g/mol |
| 8 M | Urea | 480.5 g | Urea | Mr = 60.06 g/mol |

Adjust pH to 6.3 using HCl

Denaturing Elution Buffer (1 liter):

| | | | | |
|--------|----------------------------------|---------|---|-------------------|
| 0.1 M | NaH ₂ PO ₄ | 15.6 g | NaH ₂ PO ₄ • 2 H ₂ O | Mr = 156.01 g/mol |
| 0.01 M | Tris Base | 1.2 g | Tris Base | Mr = 121.14 g/mol |
| 8 M | Urea | 480.5 g | Urea | Mr = 60.06 g/mol |

Adjust pH to 4.5 using HCl

5.2 Preparation of *E. coli* lysated under denaturing conditions

1 Cultivate and harvest cells

- *E. coli* cells may be cultivated in 96-well plates (1 mL culture volume), 24-well plates (5 mL culture volume), or any other appropriate cultivation vessel, if larger culture volumes are required.
 - Harvest cells by centrifugation at 2,000 x *g* for 15 min at 4 °C. Store cell pellets at -20 °C or -70 °C for at least 1 h.
-

2 Cell extract preparation

For preparation of cell extracts from up to 5 mL *E. coli* expression culture, we recommend the following protocol as a starting point for further optimization:

- Thaw cell pellets at room temperature. Resuspend each pellet in 0.5–1 mL of 1 x Denaturing Lysis / Equilibration Buffer. Incubate at room temperature for 30 min in a shaker.
- Centrifuge the crude lysate at 5,000 x *g* for 30 min at 4 °C to remove cellular debris. If the supernatant is not clear, centrifuge a second time to avoid clogging of the Protino® Purification Plate with insoluble material.

Note that optimal sample preparation steps have to be determined empirically depending on the characteristics of the of the polyhistidine-tagged protein and host organism.

5.3 Procedure for protein purification under denaturing conditions

Proceed to section 4.3 (vacuum manifold) or 4.4 (centrifugation) with the following modifications:

- Use Denaturing Lysis / Equilibration Buffer for the equilibration and washing step.
- Use Denaturing Elution Buffer for elution.

6 Appendix

6.1 Troubleshooting

| Problem | Possible cause and suggestions |
|--|--|
| Wells of the plate have become clogged | <i>Sample/lysate contains insoluble material</i> |
| | <ul style="list-style-type: none"> • Use centrifugation or filtration to avoid clogging. <i>Sample/lysate remains viscous from genomic DNA</i> <ul style="list-style-type: none"> • Add additional DNase |
| Protein does not bind to the resin | <i>Problems with vector construction</i> |
| | <ul style="list-style-type: none"> • Ensure that protein and tag are in frame. • Sometimes the position of the tag influences expression rate and solubility. Evaluate N- and C terminally tagged variants of the protein. • His-Tag is not accessible. • Use denaturing conditions to purify the protein. • Use a C terminal Histag instead of a N terminal tag or vice versa. |
| | <i>Incorrect binding conditions</i> |
| | <ul style="list-style-type: none"> • Check composition and pH of all buffers. Ensure that all additives are compatible (see compatibility of reagents, 2.4) |
| Protein does not elute | <i>Incorrect buffer composition</i> |
| | <ul style="list-style-type: none"> • Check composition and pH of all buffers. |
| | <i>Elution conditions are too mild.</i> |
| | <ul style="list-style-type: none"> • Increase concentration of imidazole from 250 mM to 500 mM. |
| | <i>Protein has precipitated</i> |
| | <ul style="list-style-type: none"> • Elute under denaturing conditions. |

| Problem | Possible cause and suggestions |
|--|---|
| Protein does not elute (continued) | <i>Insufficient wash</i> <ul style="list-style-type: none">• Use larger volumes for washing step.• Use NPI-50 for third washing step (containing 50 mM imidazole). |
| | <i>Binding and wash conditions are too mild</i> <ul style="list-style-type: none">• Use 10–20 mM imidazole in the binding and washing buffers. |
| | <i>Contaminating proteins and target protein are linked together via disulfide bonds</i> <ul style="list-style-type: none">• Add up to 20 mM 2-mercaptoethanol to reduce disulfide bonds. |
| Unwanted proteins elute with poly-histidine-tagged protein | <i>Contaminating proteins are proteolytic products of target protein</i> <ul style="list-style-type: none">• Perform cell lysis at 4 °C.• Include protease inhibitors. |
| | <i>Resin is not saturated with His-tagged protein</i> |
| | <i>Contaminating host proteins have a better chance to bind to the resin when only small amounts of target protein are present in the lysate. Very low amounts of polyhistidine-tagged protein are not able to replace the majority of contaminating proteins effectively.</i> <ul style="list-style-type: none">• Reduce the amount of Protino® Ni-NTA Agarose or increase the amount of sample. |
| | <i>Expression is too low</i> <ul style="list-style-type: none">• Increase expression level. Sometimes the position of the tag influences expression rate and solubility. Use a C terminal Histag instead of a N terminal tag or vice versa.• Increase amount of starting cell material. |

6.2 Ordering information

| Product | REF | Pack of |
|------------------------------|------------|--------------|
| Protino® 96 Ni-NTA | 745425.1 | 1 x 96 preps |
| | 745425.4 | 4 x 96 preps |
| Protino® Ni-NTA Agarose | 745400.25 | 25 mL |
| | 745400.100 | 100 mL |
| | 745400.500 | 500 mL |
| Protino® Ni-NTA Columns 1 mL | 745410.5 | 5 columns |
| Protino® Ni-NTA Columns 5 mL | 745415.1 | 1 column |
| | 745415.5 | 5 columns |
| Square-well Block | 740481 | 4 |
| | 740481.24 | 24 |
| Protino® Purification Plate | 745426.1 | 1 |
| | 745426.4 | 4 |
| MN Shaker Frame | 740489 | 1 |
| Square-well Block | 740481 | 4 |
| | 740481.24 | 24 |
| Self adhering PE Foil | 740676 | |
| NucleoVac Vacuum Manifold | 740681 | 1 |
| NucleoVac Vacuum Regulator | 740641 | 1 |

Visit www.mn-net.com for more detailed product information.

6.3 Product use restriction / warranty

Protino® products components are intended, developed, designed, and sold FOR RESEARCH PURPOSES ONLY, except, however, any other function of the product being expressly described in original MACHEREY-NAGEL product leaflets.

MACHEREY-NAGEL products are intended for GENERAL LABORATORY USE ONLY! MACHEREY-NAGEL products are suited for QUALIFIED PERSONNEL ONLY! MACHEREY-NAGEL products shall in any event only be used wearing adequate PROTECTIVE CLOTHING. For detailed information please refer to the respective Material Safety Data Sheet of the product! MACHEREY-NAGEL products shall exclusively be used in an ADEQUATE TEST ENVIRONMENT. MACHEREY-NAGEL does not assume any responsibility for damages due to improper application of our products in other fields of application. Application on the human body is STRICTLY

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DNA/RNA/PROTEIN purification products of MACHEREY-NAGEL are suitable for IN VITRO-USES ONLY!

ONLY MACHEREY-NAGEL products specially labeled as IVD are also suitable for IN VITRO-diagnostic use. Please pay attention to the package of the product. IN VITRO-diagnostic products are expressly marked as IVD on the packaging.

IF THERE IS NO IVD SIGN, THE PRODUCT SHALL NOT BE SUITABLE FOR IN VITRO-DIAGNOSTIC USE!

ALL OTHER PRODUCTS NOT LABELED AS IVD ARE NOT SUITED FOR ANY CLINICAL USE (INCLUDING, BUT NOT LIMITED TO DIAGNOSTIC, THERAPEUTIC AND/OR PROGNOSTIC USE).

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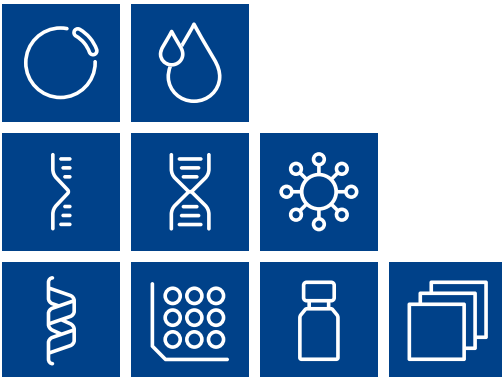
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Plasmid DNA

Clean up

RNA

DNA

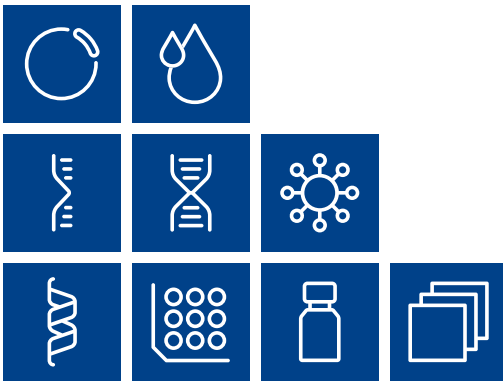
Viral RNA and DNA

Protein

High throughput

Accessories

Auxiliary tools



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