



# **Clean-up of Sequencing Reactions**

## **User Manual**

NucleoSEQ

August 2009/Rev. 02

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# 1 Components

## 1.1 Kit contents

Cat. No.	NucleoSEQ		
	10 preps 740523.10	50 preps 740523.50	250 preps 740523.250
NucleoSEQ Columns	10	50	250
Collection Tubes	10	50	250
User Manual	1	1	1

## 1.2 Material to be supplied by the user

- Collection tubes (e. g. 1.5 ml microcentrifuge tubes)

## **2 Product description**

### **2.1 The basic principle**

NucleoSEQ Columns are designed for fast and effective clean-up of nucleic acids. Using gel exclusion in a convenient spin column format allows reliable removal of smaller molecules from nucleic acids. Impurities e.g. salts, excess of labels, nucleotides, traces of organic solvents, primers are retained by the column while nucleic acids of interest are recovered with high yield. The columns are pre-filled with size exclusion matrix.

### **2.2 Kit specifications**

- Maximum sample volume to be loaded onto the column: 20 µl
- Removal of sequencing dye terminators including BigDye™

### **3 Storage conditions**

- NucleoSEQ Columns with dry gel matrix can be stored at room temperature for 12 months.
- NucleoSEQ Columns with hydrated gel matrix should be stored at 4°C. Columns can be stored up 14 days at 4°C.

### **4 Safety instructions – risk and safety phrases**

The components of the NucleoSEQ kits do not contain hazardous contents.

## 5 Protocol for sequencing reaction clean-up

### General procedure

(For details on each step see page 8.)

- 1 Spin down dried gel resin



**750 x g**  
**30 s**

- 2 Hydrate gel resin with **600 µl water**, vortex, and incubate at least **30 min** for complete hydration



**600 µl**  
**water**

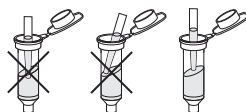
**RT**  
**> 30 min**

- 3 Remove bottom plug and spin down hydrated gel resin



**750 x g**  
**2 min**

- 4 Load sample to the center of the column



**Load**  
**column**

- 5 Spin for **4-6 min** at **750 x g** to recover purified sample



**750 x g**  
**4 - 6 min**

## Detailed procedure

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Perform sequencing reaction according to standard protocols.

We recommend to use not more than 1 -2 µl of Big Dye™ Ready reaction mix in a 20 µl sequencing reaction in order to avoid overloading the column.

---

- 1** Centrifuge the NucleoSEQ Columns for **30 s** at **750 x g** to collect the dry gel matrix on the bottom of the cartridge.
- 2** Add **600 µl dist. water** and vortex to hydrate the gel matrix. Remove air bubbles by vortexing or tapping the column. Incubate at least **30 min** or overnight to hydrate the gel matrix. Incubation can be performed at room temperature or 4°C. Hydrated columns can be stored at 4°C for a maximum of 14 days. Resuspend the settled gel matrix by inverting or vortexing the spin column several times. Air bubbles should not be visible now. Remove the bottom plug and place the spin column into a Collection Tube (supplied with the kit).
- 3** Place the column into an appropriate centrifuge (the hinge of the spin column's cover lid should be orientated to the outside of the rotor). Centrifuge **2 min** at **750 x g** to remove the remaining storage buffer. Discard the collection tube with storage buffer. Place the spin column in an appropriate collection tube (e.g. micro-centrifuge tube, not supplied with the kit).
- 4** Open the lid of the column. Carefully load the sample dropwise onto the center of the gel resin. Pipetting the sample at the sides of the spin column tube may reduce purification efficiency of the column. Moreover, do not disturb the gel surface. Sample volume should not exceed 20 µl.
- 5** Place the column in the same orientation as in step 3 into the centrifuge. Elute the sample by centrifuging the column for **4-6 min** at **750 x g**. Discard the spin column.

Dry the sample or use the sample directly.

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## 6 Appendix

### 6.1 Troubleshooting

Problem	Possible cause and suggestions
Dye blobs	<p><i>Sample added to column improperly</i></p> <ul style="list-style-type: none"> <li>Add sample directly to the center of the settled gel matrix. Dispense sample dropwise. Avoid adding sample at the sides of the gel matrix</li> </ul>
	<p><i>Sample volume to high</i></p> <ul style="list-style-type: none"> <li>Add sample in a volume of 20 µl. Higher sample volumes can cause incomplete removal of dye terminators.</li> </ul>
Poor signal intensity	<p><i>Sample volume to small</i></p> <ul style="list-style-type: none"> <li>Add sample in a volume of 20 µl. If necessary adjust sample volume to 20 µl using distilled water</li> </ul>

Conversion of RCF from different centrifuges:

$$\text{rpm} = 100 \times \sqrt{\text{RCF}/1.12r}$$

$$\text{RCF} = (\text{rpm}/1000)^2 \times 1.12r$$

rpm = revolutions per minute, RCF = relative centrifugal force ( $g$  force = RCF x  $g$ ),  
r = radius in mm

## 6.2 Ordering information

Product	Cat. No.	Pack of
NucleoSEQ	740523.10	10
NucleoSEQ	740523.50	50
NucleoSEQ	740523.250	250

Visit [www.mn-net.com](http://www.mn-net.com) for more detailed product information.

## 6.3 Product use restriction/warranty

**NucleoSEQ** kit components were developed, designed, distributed, and sold **FOR RESEARCH PURPOSES ONLY**. They are suitable **FOR IN-VITRO USES ONLY**. No claim or representation is intended for its use to identify any specific organism or for clinical use (diagnostic, prognostic, therapeutic, or blood banking).

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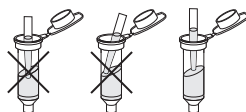
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